Brief Original Article

**In vitro antimicrobials activity against endemic *Acinetobacter baumannii* multiresistant clones**

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**Abstract**

Background: Multidrug-resistant strains of *Acinetobacter baumannii* have been reported increasingly around the world. The administration of an association of antibiotics has been proposed to create an active combination and to prevent the emergence of resistance.

Methodology: The activity of colistin, rifampicin, gentamicin, imipenem and their associations was evaluated by means of killing curves in fourteen isolates belonging to three endemic PFGE types, in a university hospital of Buenos Aires city. The 14 isolates were selected on the basis of different mechanisms responsible for resistance to carbapenems and different susceptibility to colistin.

Results: The mechanism responsible for the resistance to imipenem was the production of OXA-23 and OXA-58 carbapenemases. Heteroresistance to colistin was observed in six isolates. The associations colistin-rifampicin and colistin-imipenem were synergistic in heteroresistant isolates and prevented the development of colistin-resistant mutants. The association imipenem-gentamicin was bactericidal in gentamicin susceptible isolates, whereas the association imipenem-rifampicin was always indifferent.

Conclusion: The antimicrobial activity and the presence of synergy are related to the antimicrobials’ susceptibilities irrespective of the PFGE type or the OXA-carbapenemase produced.

**Key words:** *A. baumannii*, time-kill assay, colistin-heteroresistance, rifampicin, imipenem


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**Introduction**

*Acinetobacter baumannii* (*A. baumannii*) is an important opportunistic pathogen, particularly for patients admitted to intensive care units (ICUs); hospital outbreaks caused by this organism have increased worldwide [1-2]. The emergence of *A. baumannii* clinical isolates with resistance to multiple antimicrobials, including broad-spectrum beta (β)-lactams, carbapenems, aminoglycosides, and fluoroquinolones was observed. Carbapenem resistance has been correlated mainly with the acquisition of carbapenem hydrolysing class D β-lactamases [1]. Heteroresistance to colistin has been recently described by Li *et al.* [3].

In Argentinian hospitals, the dissemination of strains which showed epidemic behaviour and were producers of OXA-23 and OXA-58 carbapenemases has been previously documented [4-5]. For the last 15 years, our hospital has suffered a sustained endemia of multiresistant *A. baumannii* infections. The pathogen became carbapenem resistant in 1995 and since then, most of the strains have showed susceptibility to colistin and minocycline [6]. Management of *A. baumannii* multiresistant infection is a medical concern because of the limited therapeutic options available. The administration of an association of antibiotics has been proposed to create an active combination and to prevent the emergence of resistance [1].

The aim of the present study was to investigate in a university hospital of Buenos Aires city, Argentina, the activity of imipenem (IMI), colistin (COL), rifampicin (RIF) and gentamicin (GEN) alone and in double combinations against fourteen multidrug resistant *A. baumannii* isolates, producers of OXA-23 and OXA-58 carbapenemases with a high percentage of colistin heteroresistance.
Materials and methods

Fourteen clinical isolates of multidrug-resistant (MDR) *A. baumannii* were collected from routine clinical samples of patients admitted to the intensive care unit (ICU) during the period June to December 2004. Strains were identified by phenotypic tests and genomic species was determined by Amplified ribosomal DNA restriction analysis (ARDRA) [7]. Pulsed-field electrophoresis (PFGE) was performed with *ApaI* as described previously [8]. Minimal inhibitory concentrations (MICs) to COL (against the resistant subpopulations and the original strains), RIF, GEN and IMI were performed by agar dilution method and interpreted according to the Clinical and Laboratory Standards Institute (CLSI) [9] and Societe Francaise de Microbiologie for colistin and rifampicin [10]. Antibiotic powders were obtained from their respective manufacturers (COL, RIF, GEN: Sigma Chemical Co, USA; and IMI: Merck, Sharp & Dohme, Argentina). The activities of colistin, rifampicin, gentamicin, imipenem and their associations were evaluated by killing curves. The following concentrations were used: colistin 2 mg / liter; rifampicin 4 mg / liter; imipenem 8 mg / liter; gentamicin 4 mg / liter. Briefly, tubes containing cation-supplemented Mueller-Hinton broth (Lab. Britannia) with and without antibiotic were inoculated with *A. baumannii* strains in a log-phase inoculum of roughly 5x 10⁵ CFU / ml. Tubes were incubated in an ambient atmosphere at 35 °C. At time 0, 4 and 24 hours after inoculation, serial 10-fold dilutions were performed and aliquots were plated onto nutrient agar. The time-kill studies were performed twice and results were analyzed by using mean colony count values from the duplicate plates for each isolate. The bactericidal activity of single antibiotics or combinations was defined as a ≥ 3-log₁₀ CFU / ml decrease in the viable count compared with the initial inoculum. Synergism and antagonism were respectively defined as ≥ 2-log₁₀ CFU / ml decrease or increase in the viable count with the combination compared with the most active agent alone at different time points. The presence of OXA carbapenemases was searched in the fourteen selected strains by PCR assays using previously described primer pairs for the *bla_oxa23*, *bla_oxa58*, *bla_oxa40* and *bla_oxa51*-like, representative of the four OXA clusters [11]. Phenotypic screening test for metallo-ß-lactamase production using sodium mercaptoacetic acid disks was also conducted [12].

Results

Three different PFGE types were delineated in the 14 *A. baumannii* isolates. These PFGE types were identified as I, III, and IV, maintaining a previous nomenclature [4].

Twelve isolates were resistant to imipenem and only one to colistin. Rifampicin showed a high level of resistance in the majority of the isolates belonging to PFGE type I and a low level in PFGE types III and IV. All isolates belonging to PFGE type III and only one belonging to PFGE type IV were susceptible to gentamicin (Table 1). None of the isolates showed synergy in presence of sodium mercaptoacetic acid; thus they were presumed not to possess metallo-ß-lactamases [12].

In respect to the PCR results for the

| Table 1. Pharmacodynamic effects of imipenem, rifampicin and gentamicin against *A. baumannii* endemic PFGE type producers of OXA-23 and OXA-58. |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| PFGE types | N | MIC (µg/ml) | CHDL | Average changes referring to the original inoculum |
| | | IMI | RIF | GEN | IMI | CHDL | 4 hs | 24 hs | 4 hs | 24 hs | 4 hs | 24 hs | 4 hs | 24 hs | 4 hs | 24 hs |
| I | 6 | 32 | >128 | 16 | 23 | +1,2 | +2,1 | -2,0 | -3,0 | +1,4 | +2,4 | +1,0 | +3,5 | +1,2 | +2,5 | +0,5 | +3,1 |
| I | 1 | 0,5 | 2 | >16 | 8 | +1,1 | +2,0 | +2,0 | +2,1 | -1,0 | -2,3 | +1,0 | +1,1 | -1,0 | -3,1 | -0,7 | -1,8 |
| III | 3 | 32 | 8 | 0,5 | 58 | +1,1 | +2,0 | +1,1 | +1,1 | -0,7 | -1,0 | +1,1 | 0 | -0,7 | -1,8 |
| IV | 2 | 32 | 8 | >16 | 23 | 0 | +2,0 | +1,1 | +1,1 | -0,7 | -1,0 | +1,1 | 0 | -0,7 | -1,8 |
| IV | 1 | 64 | 8 | 2 | 23 | 0 | +1,6 | 0 | -1,1 | -1,0 | -1,9 | +1,2 | -0,1 | -1,0 | -4,0 |
| IV | 1 | 1 | 2 | >16 | 1 | -1,0 | -4,0 | -1,0 | -3,9 | NA | NA | -1,0 | -3,9 |

N: Number of isolates studied; CHDL: Carbapenem-hydrolysing Class D oxacillinase; NA: not assessed.
carbapenemases, the bla\textit{oxa51}like β-lactamase gene was detected in all fourteen strains whereas \textit{bla}\textsubscript{oax23} was detected in carbapenem-resistant isolates in PFGE type I and IV and \textit{bla}\textsubscript{oax58} in PFGE type III (Table 1).

Heteroresistance to colistin was observed in six isolates (Table 2).

### Table 2. Pharmacodynamic effects of colistin and colistin plus rifampicin or imipenem against \textit{A. baumannii} resistant, heteroresistant or non-heteroresistant isolates.

<table>
<thead>
<tr>
<th>PFGE types</th>
<th>(N)</th>
<th>\textit{CHDL}</th>
<th>S. col(^1)</th>
<th>MIC COL (µg/ml)</th>
<th>Average changes referring to the original ((\log_{10}CFU/ml))</th>
<th>inoculum</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>COL</td>
<td>COL-RIF</td>
<td>COL-IMI</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>4 hs</td>
<td>24hs</td>
<td>4 hs</td>
</tr>
<tr>
<td>I</td>
<td>3</td>
<td>23</td>
<td>H\textsuperscript{2}</td>
<td>1</td>
<td>-0.5</td>
<td>-3.5</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>23\textsuperscript{*}</td>
<td>no H\textsuperscript{3}</td>
<td>1</td>
<td>-2.9</td>
<td>-3.9</td>
</tr>
<tr>
<td></td>
<td>1</td>
<td>23</td>
<td>R\textsuperscript{5}</td>
<td>32</td>
<td>-1.8</td>
<td>-1.5</td>
</tr>
<tr>
<td>III</td>
<td>3</td>
<td>58</td>
<td>H</td>
<td>1</td>
<td>-1.8</td>
<td>+1.1</td>
</tr>
<tr>
<td>IV</td>
<td>4</td>
<td>23\textsuperscript{**}</td>
<td>no H</td>
<td>1</td>
<td>-3.9</td>
<td>-4.1</td>
</tr>
</tbody>
</table>

\(1\): Susceptibilities of colistin; \(2\): Heteroresistant isolates; \(3\): non-heteroresistant isolates; \(4\): resistant isolates; \(N\): Number of isolates studied. \textit{CHDL}. Carbapenem-hydrolysing Class D oxacillinase, \(*\) only in two strains, \(**\) in three strains.

### Discussion

In the present study the concentrations for the time-kill experiments were selected according to the serum-levels and the breakpoints established by CLSI, because we believe that these concentrations show better correlations with therapeutic conditions than MIC multiples in MDR strains [13-14].

In time-kill studies, imipenem showed bactericidal activity against only two carbapenem-susceptible strains, confirming its excellent activity. We observed bacteriostatic activity at four hours in all the IMI resistant strains irrespective of PFGE type or production of acquired Class D carbapenemases (Table 1). Similar results were reported by Tripodi \textit{et al}. but we believe that this behaviour would not have clinical relevance and that it would be related to the heteroresistance to carbapenemases [15].

Colistin sulphate showed bactericidal activity before 4 hs of incubation in non-heteroresistant isolates. In heteroresistant isolates we observed a decrease in the bactericidal activity or regrowth of resistant sub-populations (MIC 4-fold higher than in original strains) at 24 hours of incubation. Bacteriostatic activity was detected in the resistant isolate (Table 2). We demonstrated bactericidal activity with GEN in isolates belonging to PFGE type III and in one isolate in PFGE type IV if they were susceptible to GEN and showed synergy when combined with IMI (Table 1). The GEN concentration used is the one achieved in serum and lung tissue; hence this antimicrobial association could be a possible therapeutic option for the treatment of these MDR isolates.

In \textit{A. baumannii} infections, RIF has been used in combinations with other antimicrobials to prevent the emergence of resistant mutants. Three rifampicin MIC values were obtained: 2, 8, and > 128 µg /ml. Bactericidal activity was detected only in the isolate with an MIC of 2 ug /ml. The association IMI-RIF did not show ability to achieve synergy in isolates harbouring OXA-23 or OXA-58, because the RIF concentrations used were always lower than the MIC value (sub-MIC) (Table 1). Tripodi \textit{et al}. reported synergy with the combination IMI-RIF against producers of OXA-58 isolates using RIF concentrations higher than the MIC value in killing curves [15]. However, we observed that RIF plus COL showed synergy by inhibition of resistant sub-populations in heteroresistant isolates irrespective of the RIF MIC values and we detected synergy at 24 hours in the isolate resistant to COL despite its high MIC to both antimicrobials (Table 2). Similarly, the combination COL plus RIF was constantly synergistic against \textit{S. marcescens}, an organism that is intrinsically resistant to polymyxins [16].

In non heteroresistant isolates we were unable to demonstrate synergy between COL and IMI in isolates harbouring OXA-23 or OXA-58, partly because COL alone showed a rapid initial bactericidal effect (before four hours of incubation).
As it was previously mentioned, in heteroresistant isolates both COL-RIF and COL-IMI combinations were synergistic in inhibition of regrowth (Table 2). Similar findings were obtained with the colistin-minocycline associations [17]. Few synergy studies have been conducted in A. baumannii COL resistant strains. In agreement with a previous report by Yoon et al. [13], we detected bactericidal activity between IMI-COL or COL-RIF at 4 and 24 hours, respectively (Table 2). The rapid permeabilization of the outer membrane would enhance the action of the other antimicrobials in COL susceptible strains, whereas the synergistic mechanism in COL resistant isolates remains unknown. In accordance with Principe et al., we observed different intraclonal synergistic activities that reflect the complex interactions of multiple mechanisms of antimicrobial resistance in A. baumannii [18].

In conclusion, these studied strains belong to the three major endemic PFGE types in Buenos Aires city [4]. Thus, our results could be applied in our hospital and other medical centers for the treatment of multiresistant A. baumannii infections. From our data, we showed that the combination of COL plus RIF or IMI was effective (always bactericidal and synergistic against heteroresistant isolates) and it prevented the selection of resistance. In this study we demonstrated that the antimicrobial activity and the presence of synergy are related to the antimicrobials’ susceptibilities irrespective of the PFGE types or the OXA-carbapenemase produced.

References

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