Original Article

Detection of Salmonella human carriers in Colombian outbreak areas

Hernán Carvajal-Restrepo¹, Miryan Margot Sánchez-Jiménez¹, Sergio Díaz-Rodríguez², Nora Cardona-Castro^{1,2}

¹ Instituto Colombiano de Medicina Tropical (ICMT) – Universidad CES, Medellín, Colombia ² Facultad de Medicina Universidad CES, Medellín, Colombia

Abstract

Introduction: Salmonellosis, a zoonotic and foodborne disease, is a public health problem in developing countries. With the aim of identifying human carriers of *Salmonella*, a survey was performed in five regions of Colombia with reported salmonellosis outbreaks.

Methodology: The general population and cholecystectomy surgical patients were included in this study. Stool samples from 667 volunteers and gallbladder bile samples from 199 surgical patients were examined. Detection of *Salmonella* from cultured stool and bile samples was determined by polymerase chain reaction (PCR). Multiplex PCR and biochemical and serological tests were performed to identify the serovars of the isolates.

Results: Nine (1.35%) stool samples were positive for *Salmonella*: two *S*. Newport, two *S*. Anatum, one *S*. Sinstorf, and four *Salmonella* spp. A total of 11 gallbladder bile samples were positive: *S*. Enteritidis was isolated from 3 bile cultures (1.5%), and 8 samples (4%) were positive for *Salmonella* spp.

Conclusions: Our results show the presence of *Salmonella* carriers in the inhabitants of regions with reported outbreaks and suggest that these carriers are potential sources of infection in endemic and epidemic cases. Carriers also suggest *Salmonella* zoonotic transmission, since broiler and beef cattle are hosts to the *Salmonella* serotypes isolated. It is important to establish the source of infection in regions where salmonellosis is endemic in order to control transmission.

Key words: Salmonella; salmonellosis; carriers; zoonosis.

J Infect Dev Ctries 2017; 11(3):228-233. doi:10.3855/jidc.8023

(Received 18 December 2015 - Accepted 12 July 2016)

Copyright © 2017 Carvajal-Restrepo *et al*. This is an open-access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction

Salmonellosis, an endemic infectious disease, is considered a public health problem in developing countries. Salmonellosis is classified as a foodborne and zoonotic disease; humans acquire the infection from a variety of sources and routes of transmission [1]. However, some Salmonella serotypes differ based on geographical area, food sources (meat, vegetables, cheese, eggs, etc.), industry, and manipulation of foods [2,3]. Furthermore, the Salmonella serotype determines its ability to cause disease, as well as clinical symptoms and outbreak outcomes [4-6]. Animal and human reservoirs are responsible for the circulation of the microorganism, contributing to the occurrence of sporadic cases and outbreaks [7-10]. In addition, some food sources have higher impact on public health than do others [11-14].

Public health efforts have been made in developed countries to prevent and control foodborne and zoonotic diseases [15]. Nevertheless, the impact of these interventions is not conclusive, a fact reflected by the variability of surveillance programs for different infective sources [7]. Several mathematic and statistical approaches, such as a Bayesian model, have been developed to quantify the contributions of each major animal-food sources to human salmonellosis [15,16]. Results have identified differing sources, based on the geographic origin of the studies, that vary among eggs, pork, and layers as the most important reservoir of human salmonellosis caused in Europe by S. Enteritidis [15-17]. However, in Finland and Sweden, most cases were travel related. In most other countries, the main sources were egg-laying hens or pig reservoirs, reflecting the differences in the epidemiology of Salmonella across the European Union [18,19]. The integrated control of farms and food processing plants have shown higher reductions in the incidence of foodborne human salmonellosis [18].

Salmonellosis in Colombia is considered an endemic infection, with sporadic cases and outbreaks [5]. The global laws of food commerce recommend implementing control measures in food chains [20].

Colombia is included in these global commerce, and several guidelines are now available for producers and processers of food, such as in the poultry industry [20]. The data of salmonellosis surveillance in Colombia shows that more developed regions of Colombia report a higher number of cases each year, a fact that is not consistent with the public health infrastructure of the regions and could be interpreted as a lack of diagnosis in rural areas where diagnostic tools are unavailable [21]. Under-reporting results in registered cases of Colombian surveillance for salmonellosis, which showed that Antioquia state and the city of Bogota (the Colombian capital) contribute 33% and 29.2% of salmonellosis cases, respectively, in a report that covered the period 2000-2013 [21]. The poorest regions of Colombia, such as Chocó state, the Amazonas region, and some states on the Atlantic coast (Sucre, Magdalena, and Guajira) reported 0.1% to 0.3% of the cases [21]. The most frequent non-typhi Salmonella (NTS) reported in Colombia during the period 2000-2013 among 7,219 isolates reported were S. Typhimurium (33.7%), S. Enteritidis (28.6%), S. Dublin (3.3%), S. Derby (2.1%), S. Panamá (2.0%), S. Braenderup (1.8%), S. Saintpaul (1.7%), S. Uganda (1.4%), Salmonella spp. (1.3%), and others (24.1%, including S. Typhi 9%) [21]. The current study was carried out in Colombian regions where salmonellosis outbreaks were reported during the period of 2005-2010 [20,21], with the aim detecting human carriers of Salmonella.

Methodology

This was a cross-sectional survey to identify *Salmonella* carriers in the general population of four towns in three states located in the west, south, and east of Colombia (Quibdó, Guachené, Caloto, and Granada). Locations were chosen based on the report of salmonellosis outbreaks from 2005–2010 [20,21]. In addition, cholecystectomy surgical patients from several cities and towns of Antioquia state were included in this study (see Figure 1).

Volunteers

The general population of the above-mentioned towns were motivated to voluntarily participate in this study after a house-to-house visit by members of the research team. Participants were given individualized instructions regarding food handling and excreta disposal in the form of a brochure designed for this purpose. Instructions and a container to collect one single stool sample per volunteer were provided.

Detection of carriers in bile samples

Initially, local hospitals of the cities and towns where the stool sampling collection occurred were contacted. However, there were limitations due to the lack of surgery services at these locations. It was difficult to perform the study in rural areas. Given the difficulty of sample collection, research was conducted in two clinics in the city of Medellín that have an agreement with rural health services from Antioquia state and other Colombian regions (see Figure 1).

Sampling of bile

Patients who required a cholecystectomy because of their clinical conditions were included in the study after they signed an informed consent form. A piece of gallbladder tissue sample was stored in buffered formalin for the routine histopathology study. A separate piece of gallbladder tissue for DNA extraction was collected in 70% ethanol. The vesicular content was stored in lactose broth (Becton Dickinson, Franklin Lakes, USA).

Figure 1. Stool and bile samples studied based on the municipality of origin.



Stool and bile samples cultures

A total of 667 stool samples were collected and stored in Cary-Blair transport medium (Becton Dickinson, Franklin Lakes, USA) for culture, and in 70% ethanol for polymerase chain reaction (PCR) tests. In total, 199 bile samples were collected from patients who required a cholecystectomy.

Samples were processed at the microbiology laboratory of the Colombian Institute of Tropical Medicine (Medellin, Colombia).

Stool samples were inoculated into selenite broth (Becton Dickinson, Franklin Lakes, USA) for enrichment and incubated at 35°C for 12 hours. Then, 100 μ L of the broth was plated on MacConkey and SS (*Salmonella-Shigella*) agars (Becton Dickinson, Franklin Lakes, USA) and incubated for 12 and 24 hours, respectively, at 35°C. Additionally, the colonies grown in SS agar were plated on Rambach agar (Merck, Whitehouse Station, USA) [22].

Bile samples were collected on lactose broth, incubated at 35°C for 12 hours, then plated on blood (Becton Dickinson, Franklin Lakes, USA), MacConkey, and SS agars and incubated for 12 hours at 35°C.

For both samples, biochemical tests were performed from MacConkey agar using the API 20E

commercial kit (bioMérieux, Durham, USA) on compatible colonies.

DNA extraction

DNA extraction from bacterial colonies was made using the boiling prep method [22]. Briefly, five wellisolated colonies were seeded in 200 μ L of distilled water, shaken for 10 seconds, boiled for 10 minutes, and centrifuged at 11,000 rpm for 5 minutes [22]. The supernatant was stored in a fresh tube at -20°C until PCR was performed. DNA extraction from stool samples stored in ethanol (where the plate cultures were positive) underwent the process of extracting DNA directly using the UltraClean Fecal DNA Kit (MBio Laboratories, Carlsbad, USA). DNA extraction from bile samples in lactose broth was performed using the DNeasy Blood and Tissue Kit (Qiagen, Hilden, Germany).

PCR

All bacterial isolates, stool, and bile samples collected were tested for *hilA* gene specific to *Salmonella* following the method reported [6]. PCR conditions included 40 μ L of super mix (Invitrogen, Carlsbad, USA), 0.5 μ L of 10 μ M of forward and

Table 1. Number of bile and stool samples studied by geographical	origin.
---	---------

Department	City	Bile samples	Stool samples	Characteristics of the region	
	Medellín	106	-	6	
	Itagüí	14	-		
	Bello	12	-		
	Envigado	9	-	Urban, Andean region,	
	Copacabana	4	-	temperate climate, well to	
	Girardota	3	-	middle developed public	
	Sabaneta	2	-	health services	
	La Estrella	1	-		
	Barbosa	1	-		
	Caldas	5	-		
	Don Matías	2	-		
Antioquia	Amagá	1	-		
-	Titiribí	1	-		
	Puerto Nare	1	-	Rural, agricultural region,	
	Venecia	1	-	temperate climate, middle to	
	Santa Rosa de Osos	1	-	low developed public health	
	El Retiro	1	-	services	
	Ebéjico	1	-		
	Santafé de Antioquia	1	-		
	Marinilla	1	-		
	Apartadó	2	200	Devent the minut formet large	
	Turbo	2	-	Kural, tropical forest, low	
	Chigorodó	2	-	public health development	
Chocó	Quibdó	2	144	The second strength and the second	
Cauca	Guachené	-	204	Urban, tropical climate, poor	
Meta	Granada	-	119	public health development	
TOTAL		199	667		

reverse primers, and 9 μ L of DNA to a final volume of 50 μ L.

The primers used were [23]:

Forward: 5' - CGG AAG CTT ATT TGC GCC ATG CTG AGG TAG -3';

reverse: 5' - GCA TGG ATC CCC GCC GGC GAG ATT GTG -3'.

Amplification protocol included initial denaturation at 94°C for 5 minutes, 30 cycles at 94°C for 1 minute, 65°C for 1 minute and 72°C for 1 minute, and a final extension phase at 72°C for 10 minutes. Amplicon visualization was performed by electrophoresis in a 1% agarose gel stained with SYBR Safe (Invitrogen, Carlsbad, USA) and photographed in an image analyzer EpiChemi Dark Room (UVP, Upland, USA). The presence of an 854 bp band is a positive indicator for *hilA*. DNA from *S*. Enteritidis strain ICMT10 was used as the positive control [23].

Salmonella serotyping

All the *Salmonella* isolates were serotyped using a multiplex PCR following the established protocol [24,25]. Results of this multiplex PCR were confirmed by conventional serotyping of *Salmonella* at the reference laboratory of the National Institute of Health (Bogotá, Colombia).

Ethical considerations

This study was approved by the ethical committee of the Universidad CES. Informed consent was obtained for every participant.

Results

Samples tested

A total of 667 fecal samples and 199 bile samples were tested. The geographical origins of volunteers included in this study are shown in Figure 1 and Table 1. Briefly, 200 (30%) fecal samples analyzed were from Apartadó (Antioquia state), 144 (21.6%) from Quibdó (Chocó state), 204 (30.6%) from Guachené (Cauca state), and 119 (17.8%) from Granada (Meta state).

Stool Salmonella carriers

Nine (1.35%) stool samples were positive by culture and PCR. The distribution according serotype was as follows. Two samples from Quibdó were positive for *S*. Newport. From Apartadó, two samples were positive for *S*. Anatum, one sample was positive for *S*. Sinstorf, and one was only identified as *Salmonella* spp. In Granada, all the three isolates were identified as *Salmonella* spp. (see Table 2).

Bile Salmonella carriers

In bile samples, *S*. Enteritidis was detected by culture in three patients (1.5%) and by PCR in eight patients (4%). Table 2 shows the geographical origin of the isolates and the serotype.

By API, 34 (17.3%) bile samples were positive for bacteria: Enterobacter cloacae in - six (17.64%), Pseudomona spp. in 4 (11.76%), K. pneumoniae in four and S. Enteritidis, Pseudomonas (11.76%), fluorescens/putida, Raoultella terrigena in three (8.8%) each. For *Pseudomonas aeruginosa*, Serratia marcescens, Proteus mirabilis, Proteus penneri, Aeromonas spp., Streptococcus group A, Escherichia Enterobacter sakazakii, Stenotrophomonas coli. maltophilia, and Klebsiella oxytoca, one isolate of each was identified.

Discussion

According to the national system of surveillance and control of Colombia (National Institute of Health), from January 2008 to August 2010, 31.7% of 102 diarrheal outbreaks were due to *Salmonella* [20]. *S.* Typhimurium, *S.* Enteritidis, *S.* Branederup, *S.* Muenster, *S.* Albany, *S.* Essen, *S.* Nottingham, *S.* Give, *S.* Javiana, and *S.* Derby were the most frequent serotypes isolated in salmonellosis outbreaks from

Table 2. Salmonella isolates from stool and bile culture and/or polymerase chain reaction (PCR) in patients by geographical origin.
--

City/department	Stool culture and PCR positive	Bile culture positive	Bile PCR
Quibdó/Chocó	S. Newport (2)	Ns	Ns
	S. Anatum (2)	Ns	Ns
Apartadó/Antioquia	S. Sinstorf (1)	Ns	Ns
	S. enterica (1)	Ns	Ns
Granada/Cauca	S. enterica (3)	Ns	Ns
Guachené/Meta	(0)	Ns	Ns
Medellín/Antioquia	Ns	S. Enteritidis (2)	S. Enteritidis (2)
Envigado/Antioquia	Ns	(0)	(0)
Marinilla/Antioquia	Ns	S. Enteritidis (1)	S. Enteritidis (1)
Total	9/667	11/199	

NS: no sampling.

2005–2008. However, the source of the transmission was not determined [20]. The origins of the isolates from this study are rural and urban populations of the States of Nariño, Antioquia, Huila, Meta, and Cauca. Cities like Bogotá (the country's capital) and Medellin have high to moderate public health resources [21]. Some of these regions have the infrastructure to support the outbreaks (*i.e.*, laboratory resources and trained personnel). Other, poorer regions of Colombia do not report salmonellosis cases, likely due to their lack of laboratory and clinical resources to perform diagnoses.

S. Typhi outbreaks in Urabá (Antioquia) and in Quibdó (Chocó) were studied to determine the source of infection, notably by examining samples of residual waters in the neighborhoods where the outbreaks were reported. However, the source of outbreak was not determined [21]. Colombia is endemic for infections caused by *Salmonella*, with sporadic outbreaks [5,20,21]. Therefore, it is necessary to develop public health programs to detect carriers of *Salmonella* to reduce the risk of infection in communities.

In the current study, a systematic search for inhabiting Salmonella carriers areas where salmonellosis outbreaks occurred was performed to determine possible carriers in the affected populations. Salmonella was isolated from nine (1.35%) of 667 stool samples. From 199 bile samples, S. Enteritidis was identified by bile culture in three volunteers (1.5%). By PCR, Salmonella was detected in eight patients (4%). Projections made from the total population make the number of carriers important for salmonellosis transmission. A limitation of this study is that Salmonella serotyping was not available for five of the 8 bile/gallbladder isolates, the same limitation found in 4 of the 9 stool isolates (see Table 2).

Another limitation is the possibility that the incidence of *Salmonella* was underestimated due to the use of alternative isolation methodologies such as lactose broth used for pre-enrichment of bile samples; this has not been reported before for the culture of this kind of samples, but we used this modification to grow different types of bacteria present in bile, and this step could decrease the possibility of isolating *Salmonella*.

Public health actions, such as the active search for *Salmonella* carriers and preventive education for the control of foodborne outbreaks, are the responsibility of the governmental health institutions. However, this study shows the importance of research and academic initiatives in performing activities that influence the health and education of rural communities that might lack optimal public services and are therefore at risk of suffering foodborne outbreaks [18]. This study is

evidence of the importance of collaborative work between local government agencies and research teams to perform this type of community survey. The educational component of this work is important given that the active search for carriers is not enough to control the endemic and epidemic cases of salmonellosis in regions where the carriers were detected; an educational component may influence the behaviour of the population and therefore help to control foodborne infections. The partnership between public health agencies and research institutions identifies public health problems and develops an important educational process for the community.

Conclusions

Salmonellosis in Colombia is endemic. However, salmonellosis cases in rural regions are under-reported due to lack of laboratory infrastructures. The most frequently reported isolates in salmonellosis outbreaks that occurred between 2005 and 2008 are *S*. Typhimurium, *S*. Enteritidis, *S*. Branederup, *S*. Muenster, *S*. Albany, *S*. Essen, *S*. Nottingham, *S*. Give, *S*. Javiana, and *S*. Derby; nevertheless, the source of the transmission was not determined [20], suggesting that surveillance of salmonellosis, both as a zoonosis and as a foodborne disease, needs to be improved.

The *Salmonella* carriers identified in the same regions where the salmonellosis outbreaks were reported could be the source of transmission; however, further studies are needed in order to confirm that the isolates obtained from carriers in the present study are the same that produce the cases and outbreaks.

Acknowledgements

This work was conducted at the Instituto Colombiano de Medicina Tropical (ICMT) – Universidad CES, Medellín, Colombia. Conventional serotyping of *Salmonella* was performed at the National Institute of Health, reference laboratory (Bogotá, Colombia). Figure 1 was created by Jaime Badel. This project was financed by Colciencias grant to Nora Cardona-Castro, code 3256 408 20564.

References

- 1. Pires SM, Vieira AR, Perez E, Lo Fo Wong D, Hald T (2012) Attributing human foodborne illness to food sources and water in Latin America and the Caribbean using data from outbreak investigations. Int J Food Microbiol 152: 129-138.
- Hendriksen SW, Orsel K, Wagenaar JA, Miko A, van Duijkeren E (2004) Animal-to-human transmission of Salmonella Typhimurium DT104A variant. Emerg Infect Dis 10: 2225-2227.
- Jackson BR, Griffin PM, Cole D, Walsh KA, Chai SJ (2013) Outbreak-associated Salmonella serotypes and food commodities, United States, 1998–2008. Emerg Infect Dis 19: 1239-1244.

- 4. Pires SM, Vieira AR, Hald T, Cole D (2014) Source attribution of human salmonellosis: an overview of methods and estimates. Foodborne Pathog Dis 11: 667-676.
- Cardona-Castro N, Sánchez-Jiménez MM, Usuga-Silva LY, Arboleda-Naranjo M, Garzón E, Vélez A, Wiesner M, Muñoz N, Agudelo CI (2007) Characterization of two typhoid fever outbreaks in Apartadó, Antioquia, 2005. Biomédica 27: 236-243. [Article in Spanish.]
- Sánchez-Jiménez MM, Cardona-Castro N, Canu N, Uzzau S, Rubino S (2010) Distribution of pathogenicity islands among Colombian isolates of Salmonella. J Infect Dev Ctries 4: 555-559. doi:10.3855/jidc.670.
- Pires SM, Vigre H, Makela P, Hald T (2010) Using outbreak data for source attribution of human salmonellosis and campylobacteriosis in Europe. Foodborne Pathog Dis 7: 1351-1361.
- Craun GF (1994) Waterborne disease outbreaks in the United States of America: causes and prevention. World Health Stat Q 45: 192-199.
- Carvajal-Restrepo HN, Sánchez-Jiménez MM, Cardona-Castro NM (2010) Detection of salmonella serotypes of animal origin in inhabitants of four regions of Colombia. CES Med 24: 107-108. [Article in Spanish.]
- 10. Prouty AM, Schwesinger WH, Gunn JS (2002) Biofilm formation and interaction with the surfaces of gallstones by Salmonella spp. Infect Immun 70: 2640-2649.
- Gonzalez-Escobedo G, Gunn JS (2015) In vitro modeling of gallbladder-associated Salmonella spp. colonization. Methods Mol Biol 1225: 227-235.
- Ferrer Savall J, Bidot C, Leblanc-Maridor M, Belloc C, Touzeau S (2016) Modelling Salmonella transmission among pigs from farm to slaughterhouse: interplay between management variability and epidemiological uncertainty. Int J Food Microbiol 229: 33-43.
- 13. Im J, Nichols C, Bjerregaard-Andersen M, Sow AG, Løfberg S, Tall A, Pak GD, Aaby P, Baker S, Clemens JD, Espinoza LM, Konings F, May J, Monteiro M, Niang A, Panner U, Park SE, Schütt-Gerowitt H, Wierzba TF, Marks F, von Kalckreuth V (2016) Prevalence of Salmonella excretion in stool: a community survey in 2 sites, Guinea-Bissau and Senegal. Clin Infect Dis 62 Suppl 1: 50-55.
- 14. Corbellini LG, Júnior AB, de Freitas Costa E, Duarte AS, Albuquerque ER, Kich JD, Cardoso M, Nauta M (2016) Effect of slaughterhouse and day of sample on the probability of a pig carcass being Salmonella-positive according to the Enterobacteriaceae count in the largest Brazilian pork production region. Int J Food Microbiol 228: 58-66.
- 15. Pires SM, Hald T (2010) Assessing the differences in public health impact of salmonella subtypes using a bayesian microbial subtyping approach for source attribution. Foodborne Pathog Dis 7: 143-151.
- 16. Hald T, Vose D, Wegener HC, Koupeev T (2004) A Bayesian approach to quantify the contribution of animal-food sources to human salmonellosis. Risk Anal 24: 255-269.
- 17. Pires SM, Evers EG, van Pelt W, Ayers T, Scallan E, Angulo FJ, Havelaar A, Hald T; Med-Vet-Net Workpackage 28

Working Group (2009) Attributing the human disease burden of foodborne infections to specific sources. Foodborne Pathog Dis 6: 417-424.

- DE Knegt LV, Pires SM, Hald T (2015) Attributing foodborne salmonellosis in humans to animal reservoirs in the European Union using a multi-country stochastic model. Epidemiol Infect 143: 1175-1186.
- Wegener HC, Hald T, Lo Fo Wong D, Madsen D, Korsgaard H, Bager F, Gerner-Smidt P, Mølbak K (2003) Salmonella Control Programs in Denmark. Emer Infect Dis 9: 774-780.
- Lake R, Hudson A, Cressey P; New Zealand Food Safety Authority (2002) Risk profile Salmonella spp. (Non-typhoid) in whole chicken and in pieces. Available: http://www.foodsafety.govt.nz/elibrary/industry/Risk_Profile Salmonella-Science_Research.pdf. Accessed 24 November 2016.
- Instituto Nacional de Salud Colombia (2014) Characteristics of the isolates of Salmonella spp. Colombia. Outcome of monitoring 2000-2013. Available: http://www.ins.gov.co/tramites-y-servicios/examenes-deinter%C3%A9s-en-saludpublica/Microbiologa/Informe%20Salmonella%202000%20a %202013.pdf. Accessed 24 November 2016. [Article in Spanish.]
- Wain J, Diep TS, Bay PB, Walsh AL, Vinh H, Duong NM, Ho VA, Hien TT, Farrar J, White NJ, Parry CM, Day NP (2008) Specimens and culture media for the laboratory diagnosis of typhoid fever. J Infect Dev Ctries 2: 469-474. doi:10.3855/jidc.164.
- Pathmanathan SG, Cardona-Castro N, Sánchez-Jiménez MM, Correa-Ochoa MM, Puthucheary SD, Thong KL (2003) Simple and rapid detection of Salmonella strains by direct PCR amplification of the hilA gene. J Med Microbiol 52: 773-776.
- Muñoz Nelida, Diaz-Osorio M, Moreno J, Sanchez-Jimenez M, Cardona-Castro N (2010) Development and evaluation of a multiplex real-time polymerase chain reaction procedure to clinically type prevalent *Salmonella* serovars. J Mol Diag 12: 220-225.
- Cardona-Castro N, Sánchez-Jiménez M, Lavalett L, Múñoz N, Moreno J (2009) Development and evaluation of a multiplex polymerase chain reaction assay to identify *Salmonella* serogroups and serotypes. Diagn Microbiol Infect Dis 65: 327-330.

Corresponding author

Nora Cardona-Castro, MD, MSc, PhD Instituto Colombiano de Medicina Tropical – Universidad CES Cra. 43^a Nº 52 Sur-99 Medellín, Colombia Phone: 3053500 ext 2314 Email: ncardona@ces.edu.co

Conflict of interests: No conflict of interests is declared.