Letter to the Editor

MPT64 gene mutations leading to non-reactivity on a rapid immunochromatographic assay

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Dear Editor,

Tuberculosis is an infectious disease that remains one of the biggest killers in the world. Despite technological advances in terms of diagnostic and susceptibility testing, it still disproportionately affects communities in developing countries. Limited resources in developing countries further compound the difficulties faced when managing this illness; cost-effectiveness is a major factor to consider in determining the most suitable diagnostic tests [1]. Where resources are lacking, laboratory diagnosis may be limited to the identification of acid-fast bacilli (AFB) on direct staining. However, the sensitivity and specificity of AFB smears is limited. Ideally, facilities to culture, identify, and perform susceptibility testing of members of the Mycobacterium tuberculosis complex (MTC) are required to direct appropriate therapy. In addition, with the increase in multi-drug-resistant-tuberculosis (MDRTB) and extreme-drug-resistant-tuberculosis (XDRTB), performance of culture and susceptibility testing is becoming increasingly essential to guide treatment.

Nucleic acid based testing for tuberculosis provides a high-level of diagnostic accuracy [2]. Yet the approach is often technically complex and requires specialized equipment to perform. The high cost of such molecular assays precludes their use in resource-limited areas. Simpler, easy-to-use, rapid test alternatives are the current MPT64-based immunochromatographic (ICT) assays that are used to detect MTC from cultures. MPT64 is a MTC specific antigen secreted during bacterial growth, which has been shown to be an excellent target for differentiating MTC from mycobacteria other than tuberculosis (MOTT) [3,4]. Some M. bovis Calmette-Guérin (BCG) strains may however exhibit variable expression due to absence of the mpt64 gene [5]. Polymorphisms in the MPT64 gene have also appeared in other MTC [6]. Three commercially available MPT64-based ICT tests are: SD BIOLINE TB Ag MPT64 (Standard Diagnostics, Seoul, South Korea), BD MGIT TBc Identification Test (Becton Dickinson, Franklin Lakes, USA), and Capilia TB-Neo (Tauns, Izunokuni, Japan). Validations of these assays indicate very high sensitivity and specificity [7-11].

Our diagnostic microbiology laboratory uses the BACTEC MGIT 960 Mycobacterial Detection System (BD Diagnostics, Franklin Lakes, USA) for mycobacterial cultures. For cultures that flag positive by the MGIT system, Ziehl-Neelsen staining is performed to determine the presence of AFB with serpentine cords. When AFB with cording was identified, the SD BIOLINE TB Ag MPT64 assay was carried out according to manufacturer’s instructions. This provided a rapid identification of MTC. Between the years 2012 and 2016, eight isolates were found to be negative by the SD BIOLINE TB Ag MPT64, despite phenotypic features consistent with MTC. These isolates were later confirmed as MTC using molecular methods (Amplified MTD test (Hologic, Inc.San Diego, USA) or the Xpert TB/RIF (Cepheid, Sunnyvale, USA)), as well as an in-house PCR assay differentiating members of MTC [12]. PCR amplification and full-length gene sequencing of MPT64, including 527-base pairs upstream of the translational start site of MPT64 was performed. For six
isolates, a 63-base pair deletion between nucleotides 197 to 259 resulted in the truncation of the MPT64 protein. One isolate had a 2-base pair insertion at nucleotide 347 resulting in a frame shift mutation and another isolate had a missense mutation at nucleotide 257 (Table 1).

The 63-base pair deletion in MPT64 appears to be the most prevalent mutation followed by non-synonymous substitutions [6,13,14]. These mutations affect the structural topology and subsequently the antibody-based recognition of the MPT64 antigen by the immunochromatographic strip. The SD BIOLINE TB Ag MPT64 negative isolates in this study were not genotyped, but previous reports did not demonstrate any association between MPT64 mutations and spoligotypes [6].

Although the true prevalence of such strains is unknown, current data suggests that they are rare. Nevertheless, for laboratories that perform culture, other methods for confirmation of identification still need to be preserved. A negative MPT64 assay cannot in itself exclude the diagnosis of tuberculosis. Laboratory staff must be aware that other features such as colony morphology also need to be considered before a culture of acid-fast bacilli can be labelled as MOTT.

### References

### Table 1. Molecular characterization of mpt64 of SD BIOLINE TB Ag MPT64-negative clinical Mycobacterium tuberculosis.

<table>
<thead>
<tr>
<th>Isolate</th>
<th>Date of isolation</th>
<th>Description</th>
<th>SD BIOLINE TB Ag MPT64 result</th>
<th>MTB-BCG PCR assay*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>mpt64</td>
<td>mpt64 regulatory region</td>
<td>Effect on MPT64</td>
</tr>
<tr>
<td>TB272</td>
<td>7/1/2016</td>
<td>63bp deletion</td>
<td>WT</td>
<td>Loss of amino acids 66 to 86, leading to truncated MPT64 of 207 amino acids</td>
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<tr>
<td>TB243</td>
<td>30/11/2015</td>
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<td>WT</td>
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<tr>
<td>TB402</td>
<td>22/07/2015</td>
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<tr>
<td>TB359</td>
<td>19/01/2015</td>
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<td>WT</td>
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<tr>
<td>TB102</td>
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<td>2 bp (TC) insertion at nucleotide position 347</td>
<td>WT</td>
<td>Truncated MTP64 of 166 amino acids</td>
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<td>TB662</td>
<td>18/11/2013</td>
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<td>WT</td>
<td>Loss of amino acids 66 to 86, leading to truncated MPT64 of 207 amino acids</td>
</tr>
<tr>
<td>T278</td>
<td>10/5/2013</td>
<td>63bp deletion</td>
<td>WT</td>
<td>Loss of amino acids 66 to 86, leading to truncated MPT64 of 207 amino acids</td>
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<tr>
<td>T73</td>
<td>15/1/2012</td>
<td>Missense mutation, 257 C→G</td>
<td>WT</td>
<td>PRO86ARG</td>
</tr>
</tbody>
</table>

*WT; wild type sequence; *PCR assay for identification of M. tuberculosis complex members. (Parsons et al., 2002); ^M. tuberculosis H37Ra and M. bovis BCG Pasteur were included as positive control.


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