

# Case Report

# Extended Spectrum Beta-lactamase producing *Salmonella Lindenberg* gastroenteritis

Rajan Prabhurajan<sup>1,2</sup>, Rajasekharapanicker Kiran<sup>3</sup>, Kesavaram Padmavathy<sup>1\*</sup>

- <sup>1</sup> Research Laboratory for Oral and Systemic Health, Department of Microbiology, Sree Balaji Dental College and Hospital, Bharath Institute of Higher Education & Research (BIHER), Chennai, India
- <sup>2</sup> Department of Microbiology, Karpagam Faculty of Medical sciences & Research, Coimbatore, India
- <sup>3</sup> Salem Steel Plant Hospital, Salem, India

#### **Abstract**

Introduction: Increasing antimicrobial resistance among non-typhoidal *Salmonella* (NTS) is a major public health issue especially in developing countries and is partly due to the use of antimicrobials in animal feeds as growth promoters. NTS are often associated with self-limiting acute gastroenteritis (AGE). Nevertheless, fluoroquinolones and third-generation cephalosporins are currently used in the treatment of severe diarrhoeal infections.

Methodology: We report the case of a 30-year-old male who presented with clinical symptoms of moderate gastroenteritis. Stool culture and antibiotic susceptibility was performed as per standard microbiological methods. Molecular detection of *bla* genes was carried out by PCR. Results: The isolate was confirmed as *S. Lindenberg* by serotyping. The isolate exhibited dual resistance to fluoroquinolone and third generation cephalosporins. The isolate was an ESBL producer and harboured *blashv*. Based on the antibiotic susceptibility pattern, the patient was successfully treated with ceftriaxone-tazobactam.

Conclusion: Presently, there are no Indian reports on the *blashy* positive ESBL producing *S. Lindenberg* gastroenteritis. We report on the successful management of the first case of acute gastroenteritis caused by *S. Lindenberg* that exhibited dual resistance to fluoroquinolone and third generation cephalosporins. Continued surveillance of the antibiotic resistance pattern of the Non-typhoidal *Salmonella* serovars circulating in the geographical region is warranted.

Key words: Acute gastroenteritis; AGE; ESBL; fluoroquinolone; Salmonella; S. Lindenberg.

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### Introduction

Non-typhoidal Salmonella (NTS) serovars of the species enterica are the most common cause of foodborne salmonellosis and represent a major public health concern. NTS are often associated with selflimiting acute gastroenteritis (AGE) and are primarily transmitted by the consumption of contaminated food products of animal origin especially poultry, meat and eggs containing as few as 10<sup>3</sup> bacteria [1]. The estimated annual burden of NTS is high with the best data coming from South East Asia with a reported 22,805,000 illnesses, 37,600 deaths and an incidence per 1000 person—years of 3980 [2]. In our country, very few studies have reported the incidence of severe NTS infection but there is evidence that invasive disease is an important cause of hospital admissions in southern India, other systemic manifestations and its associated deaths [3-6].

Routine laboratory diagnosis of NTS gastroenteritis rely on the isolation of the organism by stool culture in selective media. followed by biochemical characterization and serotyping with commercially available polyvalent antisera. Though genotyping by molecular methods are available to reliably distinguish members within a serogroup, standardization, time and reagent cost limits the use of typing techniques in diagnostic clinical laboratories [7]. Whole genome sequencing is currently used as the sole, alternative method for serotype discrimination [8] but in low to middle income country settings, the isolates are generally shipped to the reference laboratories for complete serotyping. Serotyping is a skilled technique and the differentiation of closely related serotypes can take several weeks.

#### **Case Presentation**

A 30-year-old male presented to the Salem steel plant hospital, Salem, South India with history of loose stools since 2 days- 10 episodes/day, vomiting-once/day, abdominal colic, mild dehydration and low-grade fever (99.4°F). CVS: S1 S2 +, RS: Clear, Chest-no tenderness, BP: 130/84, PR: 116/minute. Ultrasound abdomen: normal study. Results from haematological tests revealed the following values: hemoglobin- 14.6 g dL $^{-1}$ , Total Count - 10,500 cells/ $\mu$ L, Platelet- 1.76 lakhs, Random blood glucose- 86 mg/dL, Urea-28 mg/dL, Creatinine-1.0 mg/dL. Serum electrolytes: Ca-9.5 mEqL $^{-1}$ , Na-134 mEqL $^{-1}$ , K-35 mEqL $^{-1}$ , Cl-104 mEqL $^{-1}$ , HCO<sub>3</sub>-26 mEqL $^{-1}$ .

Routine stool examination revealed the following, mucus 2+, Pus cells 4+, RBCs 2+, Occult bloodnegative, parasitic Ova/cysts-nil. Stool culture by standard microbiological methods yielded non-lactose fermenting gram negative bacilli. The isolate was provisionally identified as *Salmonella spp* by the standard biochemical tests. The isolate was sent to Central research institute, Kasauli, Himachal Pradesh, India for serotyping, where it was identified as *Salmonella enterica* Serovar *Lindenberg* with the antigenic structure (6,8:i:1,2).

Antibiotic susceptibility testing was performed by Kirby Bauer disc diffusion method as per CLSI guidelines [9]. The isolate was susceptible to chloramphenicol and co-trimoxazole. However, it was resistant to Ampicillin, ciprofloxacin, pefloxacin, tetracycline, ceftazidime, ceftriaxone and intermediate susceptibility was observed for cefotaxime. Nevertheless, the isolate was susceptible to ceftriaxonetazobactam. The isolate was also screened for carbapenamase production using ertapenem (10µg), meropenem (10µg), imipenem (10µg) and doripenem (10µg) discs (HiMedia Laboratories Pvt Ltd, Mumbai, India) and was found to be a non-carbapenemase producer i.e. susceptible to all the 4 carbapenems tested.

Extended spectrum beta-lactamases (ESBL) production was assessed by combined disc test (ceftazidime (CAZ: 30µg): ceftazidime-clavulanic acid (CAC: 30µg/10µg) and Cefotaxime (CTX: 30µg): cefotaxime-clavulanic acid (CEC:  $30\mu g/10\mu g$ (Himedia Laboratories Pvt Ltd, Mumbai, India). Klebsiella pneumoniae ATCC 700603 was included as the control. The test isolate exhibited an increment in zone diameter when combined with clavulanic acid (CAZ:CAC: 3 mm, CTX: CEC: 9 mm) which confirmed the production of ESBL. The minimum inhibitory concentration (MIC) was determined using Ezy MIC<sup>TM</sup> strip (CTX/CTX+CA) & Ezy MIC<sup>TM</sup> strip (CAZ/CAC+CA) (Himedia Laboratories Pvt Ltd, Mumbai, India). The MIC of Cefotaxime, cefotaximeclavulanic acid was >15µg/mL and >1µg/mL respectively, while that of ceftazidime; ceftazidimeclavulanic acid was >32µg/mL and >4µg/mL respectively indicating >3 twofold concentration decrease in MIC values when combined with clavulanate compared to MIC of the antibiotic tested alone. Further the presence of genes coding for betalactamases,  $bla_{CTX-M \text{ group 1}}$ ,  $bla_{CTX-M}$ ,  $bla_{SHV}$ ,  $bla_{TEM}$  were analysed by PCR [10-11]. BAA2146 was included as the positive control for all the 3 bla genes. The isolate harbored blashy gene nevertheless, blactx-M group 1 and blaTEM was not detected. The blaSHV PCR product had been sequenced and the sequence has been deposited in GenBank under the accession number, MK816317.

The patient was treated with IV fluids, Rantac, Rifagut, Zedott, Emeset. The patient was initially put on ofloxacin-ornidazole. Based on the antibiotic susceptibility pattern ceftriaxone-tazobactam was started. The patient responded to therapy. He was afebrile, BP: 102/70, PR: 68/min, loose stools- 2 episodes/day, vomiting- once/day, hydration-good and was discharged on the 3<sup>rd</sup> day.

#### **Discussion**

In humans, the outcome of infection with Salmonella depends primarily on the infecting serovar and the host factors [12]. NTS cause self-limiting gastroenteritis in immunocompetent patients and invasive infections in immunocompromised individuals. Among NTS, the most common serovar associated with human salmonellosis is S. enterica serovar *enteritidis* and *S. enterica* serovar *Typhimurium* [5,12]. According to the AMR surveillance network, ICMR 2017, the isolation rate of the Salmonella spp (other than S. Typhi, S. Paratyphi A and S. Typhimurium) in faeces is reported to be very high (85.7%) compared to S. Typhimurium (14.3%) [13].

Previous Indian report [14] on *S. Lindenberg* had been from invasive infections, nevertheless, here we report, for the first time a case of *S. Lindenberg* gastroenteritis. *S. Lindenberg* differs from *S. Typhimurium* in having C2 serogroup O antigens, nevertheless they possess similar H antigens, i:1,2 could lead to the misreporting of the serovar. Fluoroquinolones and third-generation cephalosporins are currently used in the treatment of severe *Salmonella* infections, while antimicrobial therapy is not indicated for mild or moderate gastroenteritis. Here, we report a case of dual fluoroquinolone resistance and ESBL production exhibited by *S. Lindenberg* that carried

*bla<sub>SHV</sub>* which is a matter of concern. This indicates that empirical treatment of AGE with ciprofloxacin /ceftriaxone could no longer be effective.

Increasing antimicrobial resistance among NTS isolates is a major public health issue and is possibly due to the use of antimicrobials in animal feeds as growth promoters. Combined ESBL and fluoroquinolone resistance among NTS has important clinical implications as this limits the antimicrobial treatment options. Hence, continued surveillance of the antibiotic resistance pattern of the NTS serovars circulating in the geographical region is warranted.

#### References

- Gupta V, Behl P (2012) Non-typhoidal salmonellosis (NTS): A global concern. J Gastrointest Infect 2: 1-2.
- Majowicz SE, Musto J, Scallan E, Angulo FJ, Kirk M, O'Brien SJ, Jones TF, Fazil A, Hoekstra RM; International Collaboration on Enteric Disease 'Burden of Illness' Studies (2010) The global burden of nontyphoidal Salmonella gastroenteritis. Clin Infect Dis 50: 882–889.
- Taneja N, Appannanavar SB, Kumar A, Varma G, Kumar Y, Mohan B, Sharma M (2014) Serotype profile and molecular characterization of antimicrobial resistance in non-typhoidal Salmonella isolated from gastroenteritis cases over nine years. J Med Microbiol 63: 66-73.
- 4. Vijaya D, Janakiram K, Sathish JV, Mohan DR, Archa S (2012) *Salmonella enteritidis* causing gastroenteritis: A case report. J Clin Diagnos Res 6: 727-728.
- Oommen S, Nair S, Nair K, Pillai S (2015) Epidemiology of non-typhoidal salmonella among patients attending a tertiary care centre in central Kerala. J Acad Clin Microbiol 17: 12-15.
- Menezes GA, Khan MA, Harish BN, Parija SC, Goessens W, Vidyalakshmi K, Baliga S, Hays JP (2010) Molecular characterization of antimicrobial resistance in non-typhoidal salmonellae associated with systemic manifestations from India. J Med Microbiol 59: 1477–1483.
- Alvarez J, Sota M, Vivanco AB, Perales I, Cisterna R, Rementeria A, Garaizar J (2004) Development of a multiplex PCR technique for detection and epidemiological typing of

- Salmonella in human clinical samples. J Clin Microbiol 42: 1734-1738.
- 8. Ibrahim GM, Morin PM (2018) *Salmonella* serotyping using whole genome sequencing Front Microbiol 13: 2993.
- Cinical and Laboratory Standards Institute (CLSI) (2018)
   Performance standards for antimicrobial susceptibility testing:
   Twenty Eighth International Supplement, CLSI document
   M100-S28 (ISBN 1-56238-839-8).
- Woodford N, Fagan E J, Ellington MJ (2006) Multiplex PCR for rapid detection of genes encoding CTX-M extendedspectrum β-lactamases. J Antimicrob Chemother 57: 154–155.
- Hassan M, Alkharsah K, Alzahrani A, Obeid O, Khamis A, Diab A (2013) Detection of extended spectrum betalactamases-producing isolates and effect of AmpC overlapping. J Infect Dev Ctries 7: 618-629. DOI: 10.3855/jidc.2919.
- World Health Organisation (2018) Fact-sheets Salmonella-(non-typhoidal) Available: www.who.int/news-room-fact-sheets/detail/salmonella-(non-typhoidal) Accessed: 20 February 2019.
- Indian Council of Medical Research (2017) Annual report -Antimicrobial Resistance Surveillance Network January 2017-December 2017. Available: https://www.icmr.nic.in/sites/default/files/reports/annual\_report\_amr\_jan2017-18.pdf. Accessed: 23 April 2018.
- Sudhaharan S, Kanne P, Vemu L, Bhaskara A (2018) Extraintestinal infections caused by nontyphoidal *Salmonella* from a tertiary care center in India. J Lab Physicians 10: 401-405.

## **Corresponding author**

Kesavaram Padmavathy, Ph,D
Associate Professor, Department of Microbiology,
Research Laboratory for Oral and Systemic Health,
Sree Balaji Dental College and Hospital,
Bharath Institute of Higher Education & Research (BIHER),

Chennai, India, Pincode: 600100.

Tel: +91-9884164212

Email: padmabakianath@gmail.com

**Conflict of interests:** No conflict of interests is declared.