

Original Article

Prediction of sofosbuvir response using interleukin-6 serum level and single nucleotide polymorphism of interferon lambda-4Amal E Saafan¹, Ashraf Abobaker², Mohamed S Abbas², Ahmed O El-Gendy³¹ Department of Pharmaceutical Microbiology, Faculty of Pharmacy, Menoufia University, Shebin ElKoum, Egypt² Military Medical Academy, Cairo, Egypt³ Microbiology and Immunology Department, Faculty of Pharmacy, Beni-Suef University, Egypt**Abstract**

Introduction: In Egypt, 15% of the populations are suffering from chronic hepatitis C especially genotype 4. Sofosbuvir was approved by FDA in December 2013 for treatment of HCV genotypes 2 and 3 in combination with Ribavirin, and for genotypes 1 and 4 in combination with Peg-IFN. Recently, polymorphism of different genes and plasma levels of IL-6 were utilized for better prediction of HCV clearance. This study aimed at early prediction of the efficacy of HCV treatment with Sofosbuvir (Sovaldi) and comparing the antiviral efficacy of dual and triple Sovaldi combination therapy.

Methodology: Blood samples were collected from 100 HCV positive patients and detected by real time PCR at three time intervals. SNP genotyping of INFL-4 gene was estimated by using real-time PCR with predesigned primers and Taqman probes. IL-6 serum level was estimated before, during and after the end of the treatment using ELISA assay based on human IL-6 KIT.

Results: SNP genotyping of INFL-4 gene showed that 13.1% of patients carried $\Delta G/\Delta G$, 30.4% patients had TT/TT and 56.5% patients possessed heterozygote allele $\Delta G/TT$. Clinical data displayed that 13 patients were got relapsed at SVR 12. Serum level of IL-6 was noticed higher in HCV patients than healthy ones. Noteworthy, it was increased during treatment then decreased to a minimal level than beginning of treatment.

Conclusion: SNP in INFL-4 gene has displayed no effect in response to Sofosbuvir. Dual therapy had the same effect like triple therapy, so interferon could be withdrawn from the treatment regimen.

Key words: Sofosbuvir; IL6; IFNL4; HCV.*J Infect Dev Ctries* 2020; 14(1):80-88. doi:10.3855/jidc.12013

(Received 11 September 2019 – Accepted 07 December 2019)

Copyright © 2020 Saafan *et al.* This is an open-access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.**Introduction**

Egypt suffers from a particular high morbidity and mortality rate with 40,000 patients who were dying from hepatitis C viral (HCV) infection each year. It was found that nearly 15 million Egyptians are currently suffering from this virus. Every year, there are 170,000 to 200,000 new HCV cases [1]. Being of the high prevalence in Egypt is due to needles reuse during mass-treatment programs for schistosomiasis at the end of the 20th century. Till now, transmission continues to occur, primarily through iatrogenic sources, such as blood transfusions, injections, and dental care [2,3]. HCV is a positive sense strand RNA virus, which characterized by a high heterogeneity of sequence. There are seven HCV genotypes 1 to 7 with a large number of subtypes [4,5], and the major genotype in Egypt is genotype-4 [6]. Sofosbuvir as nucleotide polymerase inhibitor that was developed as an oral drug for the treatment of chronic HCV infection [7]. It acts

through phosphorylation within the host hepatocyte to the active Sofosbuvir triphosphate, which competes with the natural nucleotides, thereby causing termination of RNA replication in the nascent viral genome. The active triphosphate of Sofosbuvir targets the highly conserved active site of the HCV-specific NS5B (nonstructural 5B) polymerase, acting as a non-obligate chain terminator, an effect that is independent of the viral genotype [8,9]. Regimen of Sofosbuvir combination in the treatment of HCV may undergo first-dual regimen in which Sofosbuvir (400 mg/day) and Ribavirin (1000–1200 mg/day according to weight) [10] used for 24 weeks. In addition, second–triple regimen Sofosbuvir (400 mg/day) in combination with peg-interferon (1200 mg/week) and Ribavirin (1000–1200 mg/day) for 12 weeks possibly to achieve a high sustained virological response “SVR” [11]. The interferon-lambda-4 (IFNL4) gene was discovered to have a functional dinucleotide variant (rs368234815,

previously designated as ss469415590; IFNL4-TT/ Δ G) that was located within the first IFNL4 exon. IFNL4- Δ G genotype predicts slower viral decline and lower odds of SVR to IFN- α /RBV treatment better than IL28B (rs12979860) because the deletion frame shift IFNL4- Δ G allele creates an open reading frame that allows production of a novel IFN- λ 4 which causes over expression of IFN-stimulated gene (ISG) in genotype-1 [12]. IFN-L4 polymorphism seems to be the best single predictor of SVR in genotype 3 infected patients [13]. Interleukin-6 (IL-6) is pleiotropic cytokine that has an important role in an acute phase response [14]. IL-6 was produced in the liver by kupffer cells in response to HCV infection [15] and induced the production of C-reactive protein [16]. IL-6 was increased in patients with liver disease such as a chronic hepatitis C or HBV when compared to healthy subjects [17,18]. Hepatitis C causes toll like receptor expression and increase IL-6 secretion by human B cells in vitro [19]. It was shown that IL-6 is correlated with resistance to Pegylated interferon/Ribavirin (PEG-INF/RBF) treatment [20].

Herein, the relation between serum level of IL-6 and resistance to treatment of HCV genotype-4 by a newly approved direct acting antiviral treatment Sofosbuvir has been investigated. Also, the role of INF-L4 polymorphism and serum level of interleukin-6 as a predictor factor in the treatment of HCV genotype -4 by Sofosbuvir has been presented. Till now, there is no study in Egypt about Sofosbuvir and INF-L4 polymorphism and that compares efficacy of dual and triple regimen.

Methodology

Ethical considerations

Informed consent was obtained from all subjects before collection of blood samples. All procedures were approved by the ethics committee of Faculty of Pharmacy, Beni-Suef University (23/2015).

Patient recruitment and exclusion criteria

One hundred patients, who chronically infected with HCV genotype 4 from different Egyptian healthy centers, were eligible for enrollment. Blood samples were collected from patients in the period from January 2015 till April 2015. Only 92 patients continued till the end of the study (20 patients had administered triple therapy and 72 had administered dual therapy). The remaining 8 patients stopped the treatment due to the encountered severe side effects. The control of our study was 8 normal persons. Fibrosis stages were determined by fibro scan according to the Metavir scoring system F1 –4.

Exclusion criteria: ascites or history of ascites, hepatic encephalopathy or history of hepatic encephalopathy and serum creatinine > 2.5 mg/dL. If creatinine was between 1.5 and 2.5 md/dL, an estimated glomerular filtration rate (eGFR) should be calculated and should be exceeded 30 mL/minute with favorable nephrological consultation. Other exclusions included extra-hepatic malignancy except after two years of disease-free interval; pregnancy or inability to use effective contraception; cardiac patients who were receiving amiodarone as treatment; hemoglobin < 10 mg/dL; platelets < 50,000/mm; HBsAg positive and if patients provided data showing HCV genotype other than genotype 4.

Blood sampling

Blood sample was collected from all patients before starting the treatment and then divided into two parts; one in EDTA tube for DNA extraction and single nucleotide polymorphism (SNPs) genotyping and second part in a plane tube centrifuged to separate serum and kept in -80° C refrigerator. Also, blood samples were collected at the 4th week after starting treatment and at the end of treatment in order to investigate for IL-6 levels.

DNA extraction

Blood samples in EDTA were used for DNA extraction manually by using; G-spin total DNA extraction kit from (Intron Biotechnology, Korea) according to manufacturer instructions DNA purity and concentration were calculated by using a Nano drop (Clinilab Cairo, Egypt).

SNP Genotyping for INF-L4 at ss469415590

A Taqman probe assay was conducted using a predesigned primers & probes purchased from Integrated DNA Technology (IDT) Company, USA. The used primers were the forward primer: ss469415590IFNL4_F GCCTGCTGCAG AAGCAGAGAT and the reverse primer: ss469415590IFNL4_R GCTCCAGCGAGCGGTAGTG. Also, the probes were ss469415590IFNL4_HEX ATCGCAGAAGGCC-BHQ and ss469415590IFNL4_MFAM-ATCGCAGCGGCC-BHQ [12].

All reactions were set up using 1 μ l isolated gDNA and TaqMan Genotyping Master Mix at which genotyping was done by a Step One Plus instrument (Life Technology, Carlsbad, California, USA) performed in Clinilab Cairo, Egypt. Temperature

cycling profiles were as follows; an initial denaturation at 95°C for 10 minutes, followed by 40 cycles of denaturation of DNA at 95°C for 10 seconds, annealing of primers at 58°C for 5 seconds and extension at 72°C for 20 seconds. The detection of fluorescent products was monitored once every cycle.

Quantitative determination of IL-6 serum level

In order to investigate for IL-6 levels in serum samples, an ELISA assay based on human IL-6 kit (Sun Red Biotechnology Company, Shanghai, China) was used according to manufacture instruction.

Quantitative estimation of HCV- RNA using real-time PCR

Blood samples after four weeks from starting the treatment were taken to determine rapid virological response (RVR), then samples were taken at the end of the treatment to determine end of the treatment virological response (ETVR). Also, samples were collected after twelve weeks from the end of the treatment to determine sustained virological response-12 (SVR12). Lastly, samples were taken after six months from the end of the treatment to determine sustained virological response-24 (SVR24).

In all cases, viral RNA was extracted from blood plasma using RNeasy Spin-Columns ZYMO RESEARCH"ZR" Viral RNA Kit™ (Catalog No. R1034) provided for rapid isolation of high-quality viral RNA from biological sources. For quality reason, an internal control sample (IC) corresponding to a stabilized RNA fragment was added to each sample prior to RNA extraction. The HCV quantitative real-time PCR was conducted using one step HCV quantitative Taqman probes based kit (IVD, DNA-Technology, Research & Production, LLC, Russia) according to manufacturer instructions. RNA reverse transcription step was done by incubating the reaction tube at 40 °C for 30 minutes and then incubated at 95 °C for 5 minutes. DNA probes used for nucleic acid (NA) and internal control (IC) PCR products detection were labeled with FAM and HEX fluorescent probes respectively. The amplification process consisted of an initial denaturation at 94 °C for 5 minutes the a 50 repeated cycles of: thermal DNA denaturing at 94 °C for 10 seconds, primer annealing with complementary sequences and further polynucleotide chains completion by Taq-polymerase at 62 °C for 20 seconds including the optical measurement at this step. The quantitation of the HCV RNA was performed with an aid of four standards (STs) with known concentration of artificially synthesized target DNA. The STs were

used to build the standard curve which was necessary to quantitate the RNA in each sample.

Statistical analysis

Data were coded and entered using the statistical package SPSS (Statistical Package for the Social Sciences) version 25. Data was summarized using mean, standard deviation, median, minimum and maximum in quantitative data and using frequency (count) and relative frequency (percentage) for categorical data. Comparisons between quantitative variables were done using the non-parametric Kruskal-Wallis and Mann-Whitney tests [21]. For comparing categorical data, Chi-squared test was performed. Exact test was used instead when the expected frequency is less than 5 [22]. Receiver Operating Characteristic curve was constructed with area under curve analysis performed to detect best cutoff value of IL-6 for detection of response. P-values < 0.05 were considered as statistically significant.

Results

A number of 92 HCV patients with a mean age 53 years and a number of 8 healthy controls with a mean age of 42 years were enrolled in this study. About 73.91% (n = 68) of cases were females and 26.09% (n = 24) were males, while the distributions of controls were 50% males and 50% females.

Virological characteristics of patients infected with HCV after dual and triple Sovaldi combination therapy

The overall results after dual or triple therapy in terms of rapid virological response (RVR) showed that 19.6% (n = 18) were positive for HCV by PCR, while 80.4% (n = 74) were negative. While after the end of treatment by 12 weeks (SVR12), 17.4% (n = 16) were positive and 82.6% (n = 76) cases were negative. Regarding SVR24, there was no difference compared to SVR12. The detailed virological characteristics of dual and triple Sovaldi combination therapy are well-illustrated in Table 1. There was no statistically significant difference between both groups regarding RVR, SVR12, and SVR24, as shown in Table 1.

Frequencies of interferon lambda-4 genotypes and their association with different virological response

The frequency of percentages of IFN-L4 SNP genotypes whether it was TT/TT, TT/G or G/G alleles in different patients and control cohorts are illustrated in Figure 1.

Figure 1. Frequencies of interferon-lambda genotypes in different patients and control cohorts.

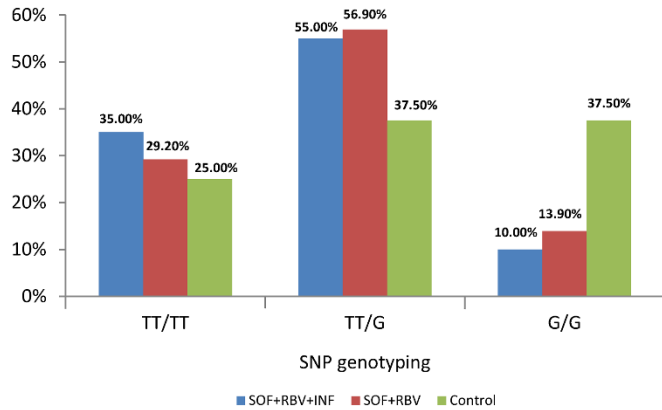


Figure 2. Comparison of serum levels of interleukin-6 before starting therapy.

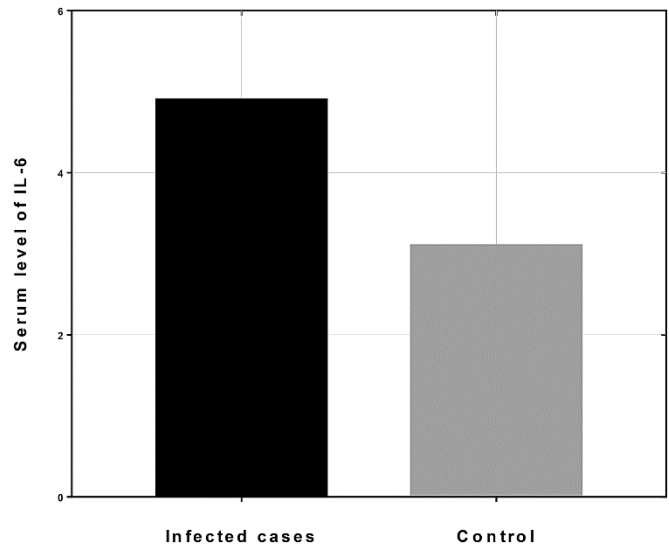


Table 1. Comparison between the antiviral efficacy of dual and triple Sovaldi combination therapy.

		Treatment plan				P value
		Triple therapy group		Dual therapy group		
		Count	%	Count	%	
RVR	Negative	17	85.0%	57	79.2%	0.75
	Positive	3	15.0%	15	20.8%	
SVR12	Negative	17	85.0%	59	81.9%	1
	Positive	3	15.0%	13	18.1%	
SVR24	Negative	17	85.0%	59	81.9%	1
	Positive	3	15.0%	13	18.1%	

RVR: rapid virological response; SVR12: sustained virological response-12; SVR24: sustained virological response-24.

Table 2. Relation of SVR12 with frequencies of IFN-L4 genotypes (TT/TT, TT/G, G/G).

		SVR12				P value	
		Negative		Positive			
		Number	%	Number	%		
Triple therapy group	SNP genotyping	TT/TT	6	35.3%	1	33.3%	0.32
		TT/G	10	58.8%	1	33.3%	
		G/G	1	5.9%	1	33.3%	
Dual therapy group	SNP genotyping	TT/TT	16	27.1%	5	38.5%	0.62
		TT/G	35	59.3%	6	46.2%	
		G/G	8	13.6%	2	15.4%	
Total	SNP genotyping	TT/TT	22	28.9%	6	37.5%	0.46
		TT/G	45	59.2%	7	43.8%	
		G/G	9	11.8%	3	18.8%	

SNP: single nucleotide polymorphism.

Frequencies of IFN-L4 genotypes (TT/TT, TT/G, G/G) in HCV infected patients and different treatment outcomes

Of the negative SVR12 cases, 28.9% (n = 22) were TT/TT SNP genotyping, 59.2% (n = 45) were TT/G, and 11.8% (n = 9) were G/G. Of the positive SVR12 cases, 37.5% (n = 6) had TT/TT SNP genotyping, 43.8% (n = 7) had TT/G, and 18.8% (n = 3) had G/G. There was no significant difference between SVR12 positive and negative cases regarding various SNP genotyping, with p-value = 0.46, as shown in Table 2.

Frequencies of IFN-L4 genotypes (TT/TT, TT/G, G/G) in triple therapy group with different treatment outcomes

In negative SVR12 cases, 35.3% (n = 6) had TT/TT SNP, 58.8% (n = 10) had TT/G, and 5.9% (n = 1) had G/G. In positive SVR12 cases, 33.3% (n = 1) had TT/TT SNP, 33.3% (n = 1) had TT/G, and 33.3% (n = 1) had G/G. There was no significant difference between SVR12 negative and positive cases regarding SNP genotyping in triple therapy group.as shown in Table 2.

Frequencies of IFN-L4 genotypes (TT/TT, TT/G, G/G) in dual therapy group and different treatment outcomes

In negative SVR12 cases, 27.1% (n = 16) had TT/TT, 59.3% (n = 35) had TT/G, and 13.6% (n = 8) had G/G. In positive SVR12, 38.5% (n = 5) had TT/TT, 46.2% (n = 6) had TT/G, and 15.4% (n = 2) had G/G, with no significant difference as shown in Table 2.

Interleukin-6 serum levels

Interleukin-6 level was determined by ELISA assay kit. The mean level of interleukin-6 was higher in infected cases (5.5 ± 2.2) than in healthy control, with a

mean level of 3.1 ± 0.18 that showed a significant difference (p = 0.002) as shown in Table 3 and Figure 2.

Association of interleukin-6 with virological response Levels of interleukin-6 in HCV infected patients in relation to treatment

Serum level of IL-6 before treatment in SVR12 negative cases ranged from 1.8 to 10.5 pg/mL, with mean value of 5.5 ± 2.1 pg/mL. On the other hand, in SVR12 positive cases, it ranged from 2.2 to 10.0 pg/mL, with mean value of 5.4 ± 2.5 pg/mL. Serum level of IL-6 during treatment in SVR12 negative cases ranged from 7.5 to 11.1 pg/mL, with mean value of 9.8 ± 0.8 pg/mL, but in SVR12 positive cases, it ranged from 8.5 to 10.8 pg/mL, with mean value of 10.0 ± 0.7 pg/mL. Serum level of interleukin-6 after treatment in SVR12 negative cases ranged from 1.2 to 10.7 pg/mL, with mean value of 4.2 ± 1.9 pg/mL, while in SVR12 positive cases, it ranged from 3.2 to 8.1 pg/mL, with mean value of 5.9 ± 1.4 pg/mL. There were no statistically significant differences between SVR12 positive and negative cases regarding serum levels of interleukin-6 before and during treatment, with p-value 0.7, and 0.4, respectively. However, serum level of IL-6 was significantly higher in SVR12 positive cases than SVR12 negative cases, after treatment, with p-value 0.001 as shown in Table 4.

Levels of interleukin-6 in triple therapy group in relation to treatment

In triple therapy group; before treatment, serum level of interleukin-6 in SVR12 negative cases ranged from 2.5 to 8.7 pg/mL, with mean value of 5.4 ± 2.2 pg/mL, and in SVR12 positive cases, it ranged from 3.6 to 9.6 pg/mL, with mean value of 6.0 ± 3.2 pg/mL.

Table 3. Comparison of serum levels of interleukin-6 before starting therapy.

	Infected cases				Control				p-value
	Mean ± SD	median	Min	Max	Mean ± SD	median	Min	Max	
Serum level of IL-6	5.5±2.2	4.9	2.5	10.5	3.1±0.18	3.1	2.9	3.5	0.002

IL-6: Interleukin-6.

Table 4. Levels of interleukin-6 in all patients in relation to treatment.

Serum level of IL-6	SVR12										P value
	Negative					Positive					
	Mean	SD	Median	Min	Max	Mean	SD	Median	Min	Max	
Before	5.5	2.1	4.9	1.8	10.5	5.4	2.5	5.2	2.2	10.0	0.71
During	9.8	0.8	9.9	7.5	11.1	10.0	0.6	10.0	8.5	10.8	0.39
After	4.2	1.9	3.8	1.2	10.7	5.9	1.4	6.1	3.2	8.1	0.001

IL-6: Interleukin-6; SVR12: sustained virological response-12.

Figure 3. ROC curve for prediction of response using IL-6 in triple therapy.

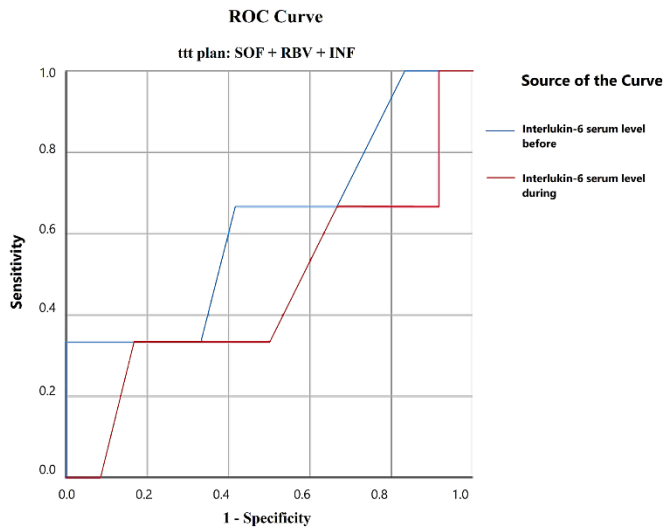


Figure 4. ROC curve for prediction of response using IL-6 in dual therapy.

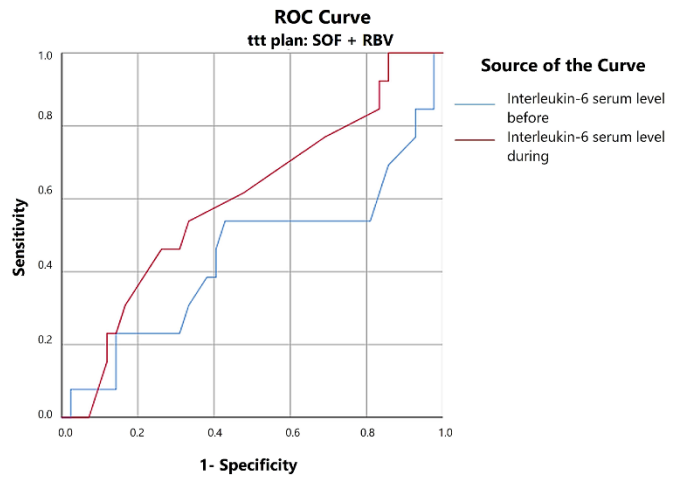


Table 5. Levels of interleukin-6 in triple and dual therapy group in relation to treatment outcome.

Interleukin-6 serum level		SVR12									P value	
		Mean	SD	Negative Media n			Positive Media n					
				Min	Max	Mean	SD	Min	Max			
Triple therapy	Before	5.4	2.2	4.7	2.5	8.7	6.0	3.2	4.9	3.6	9.6	0.54
	During	10.0	0.7	10.0	8.3	10.9	9.6	1.0	9.9	8.5	10.5	0.84
	After	4.0	2.1	3.7	1.2	7.2	6.9	0.4	7.1	6.5	7.1	0.07
Dual therapy	Before	5.5	2.1	5.0	1.8	10.5	5.2	2.4	5.5	2.2	10.0	0.53
	During	9.8	0.8	9.8	7.5	11.1	10.1	0.6	10.0	8.9	10.8	0.21
	After	4.2	1.8	3.8	1.2	10.7	5.7	1.5	6.0	3.2	8.1	0.01

SVR12: sustained virological response-12.

Table 6. Prediction of response to triple and dual therapy using IL-6.

Type of the therapy	Time	Area Under the Curve	P value	95% Confidence Interval	
				Lower Bound	Upper Bound
Triple therapy	Interleukin- 6 serum level before treatment	0.63	0.52	0.24	1.00
	Interleukin- 6 serum level during treatment	0.458	0.829	0.051	0.87
Dual therapy	Interleukin-6 serum level before treatment	0.44	0.53	0.24	0.64
	Interleukin- 6 serum level during treatment	0.6	0.29	0.42	0.77

During treatment, in SVR12 negative cases, it ranged from 8.3 to 10.9 pg/mL, with mean value of 10.0 ± 0.8 pg/mL, but in SVR12 positive cases, it ranged from 8.5 to 10.5 pg/mL, with mean value of 9.7 ± 1.0 pg/mL. After treatment, in SVR12 negative cases, it ranged from 1.2 to 7.2 pg/mL, with mean value of 4.0 ± 2.1 pg/mL, while in SVR12 positive cases, it ranged from 6.5 to 7.1 pg/mL, with mean value of 6.9 ± 0.35 pg/mL. There were no statistically significant differences between SVR12 positive and negative cases regarding serum levels of IL-6 before, during, and after treatment, with p-value 0.54, 0.84, and 0.07, respectively as shown in Table 5.

Receiver Operating Characteristic (ROC) curve for prediction of response using IL-6 in triple therapy

Area under the curve for IL-6 serum level before triple therapy was 0.625, with 95% confidence interval (0.238-1.000), and p-value 0.516, while during therapy, it was 0.458, with 95% confidence interval (0.051-0.865), and p-value 0.829 shown in Table 6 and Figure 3.

Levels of interleukin-6 in dual therapy group in relation to treatment

In dual therapy group; before treatment, serum level of interleukin-6 in SVR12 negative cases ranged from 1.8 to 10.5 pg/mL, with mean value of 5.5 ± 2.1 pg/mL, and in SVR12 positive cases, it ranged from 2.2 to 10.0 pg/mL, with mean value of 5.2 ± 2.4 pg/mL. During treatment, in SVR12 negative cases, it ranged from 7.5 to 11.1 pg/mL, with mean value of 9.8 ± 0.8 pg/mL, but in SVR12 positive cases, it ranged from 8.9 to 10.8 pg/mL, with mean value of 10.1 ± 0.6 pg/mL. After treatment, in SVR12 negative cases, it ranged from 1.2 to 10.7 pg/mL, with mean value of 4.3 ± 1.8 pg/mL, while in SVR12 positive cases, it ranged from 3.2 to 8.1 pg/mL, with mean value of 5.7 ± 1.5 pg/mL. There were no statistically significant differences between SVR12 positive and negative cases regarding serum levels of interleukin-6 before, and during treatment, with p-value 0.53 and 0.29 respectively. There was statistically significant difference between SVR12 positive and negative cases regarding serum levels of interleukin-6 after treatment, with p-value 0.01 as shown in Table 5.

ROC curve for prediction of response using IL-6 in dual therapy

Area under the curve (AUC) for interleukin-6 serum level before dual therapy was 0.442, with 95% confidence interval (0.24-0.64), and p-value 0.53, while during therapy, AUC was 0.597, with 95% confidence

interval (0.42-0.77), and p-value 0.29 as shown in Table 6 and Figure 4.

Discussion

The highest prevalence in the world of HCV infection in Egypt is (18%) [23], so the treatment of HCV in Egypt is the national project. Firstly, HCV was treated with pegylated-interferon (PEG-IFN) plus Ribavirin [24] but this combination provided low rate of SVR about 40% to 50% and was accompanied by some intolerable side effects [25]. Sofosbuvir was the first drug which used orally once daily and provide SVR exceed 90% [26]. In this study, we selected 100 patients according to exclusion and inclusion criteria that have been mentioned above and 8 patients didn't complete to the end of the study.

Both SVR12 and SVR24 were the same at which sixteen cases (17.4%) were positive, and 76 (82.6%) cases were negative as shown in Table 1. It was found a concordance between SVR12 and SVR24 measurements [27,28]. In contrast, to our results shown here, that based on HCV genotype-4 patients, Thorlund and his coworkers observed that there was a big difference between SVR12 and SVR24 who studied hepatitis C genotype 1 patients treated with peg-interferon plus Ribavirin [29].

Furthermore, there were no statistical significant differences between triple therapy regimen and dual therapy of RVR, SVR12, and SVR24 as shown in Table 1. These results were in agreement with that reported by Satsangi, *et al.*, [30] who showed in their study 13 cases infected with HCV genotype-4 and received dual and triple therapy regimen fulfilling SVR12 identical in both regimens, so the deletion of peg-interferon is useful to avoid side effects and economic costs of peg-interferon.

Further, it was shown that there was no significant difference between SVR12 positive and negative cases regarding to various SNPs genotyping of IFN-L4, with p-value = 0.46, as shown in Table 2. This means that patient with IFN-L4 $\Delta G/\Delta G$ genotypes had a lower percentage in achieving SVR than those carry TT/ ΔG or TT/TT genotypes but these differences were not significant statistically. These results have boozed the study done by Meissner and coworkers [12] and are in agreement with Stättermayer and his group [31]. However, Stättermayer study was statistically significant on HCV patients treated by Ribavirin and interferon only. But our study and Meissner's study included patients treated by DAA drugs. These statistically difference in three studies may be due to the number of samples.

The serum level of interleukin-6 before treatment has been found significantly higher in HCV patients than in healthy control as shown in Table 3. This finding was in agreement with the result of El Salam, [32] and ElSerafi group [33]. These came with the result of study which done by Feldman *et.al* [34] who proved that HCV infection stimulate toll like receptor -2 by HCV core protein which resulted in induction of interleukin-6 and other inflammatory cytokines secretion.

Also, the serum level of interleukin-6 before treatment was slightly higher in the group that achieved SVR than others who did not achieve SVR as shown in Table 4. On the other hand, there was no significant difference between the two groups and this was confirmed by the ROC curve that was used to study the sensitivity and specificity of serum level of IL-6 in predicting the response of treatment as illustrated in Table 6 and figures 3, 4. This agrees with the study which was done by Cotler [35] who stated that there was no correlation between SVR and baseline of serum levels of interleukin-6. In contrast to, Nattermann [36] and Faisal [37] who stated that serum level of IL-6 was higher in group who achieved SVR than those who did not and this could be used as a response predictor.

Eventually, serum level of IL-6 was increased significantly during treatment, then after the end of the treatment, the level of IL-6 was decreased significantly as shown in Table 5. These came in line with Villani [38] and Tarragô [39] who stated that IL-6 level was increased at the beginning of the treatment then decline to reach the level less than that of the pre-treatment. This was explained as, the eradication of virus by Sofosbuvir treatment regimens lead to restoration of immune response. In contrast to this study which done by Saraiva [40] who stated that serum level of interleukin-6 didn't change after the treatment due to its role in liver repair process.

Conclusion

SNP of INFL-4 gene has no effect in response to Sofosbuvir and SVR12 or SVR24 showed similar results but a further follow up at SVR12 is recommended. Dual therapy had the same effects as triple one so; changes in treatment regimen based on the data presented herein should be reconsidered by withdrawing interferon from the treatment regimen. IL-6 level increased at the start of the treatment then decreased till reach to the level less than pretreatment.

Acknowledgements

Dr. Amir Ibrahim and Dr. Bakheet Elkot are specially acknowledged for their support and technical assistance.

References

1. Elgharably A, Gomaa AI, Crossey MM, Norsworthy PJ, Waked I., Taylor-Robinson SD (2017) Hepatitis C in Egypt—past, present, and future. *Int J Gen Med* 10: 1-6.
2. Strickland GT (2006) Liver disease in Egypt: hepatitis C superseded schistosomiasis as a result of iatrogenic and biological factors. *Hepatology* 43: 915-922.
3. Buckley GJ, Strom BL (2017) A national strategy for the elimination of viral hepatitis emphasizes prevention, screening, and universal treatment of hepatitis C. *Ann Intern Med* 166: 895-896.
4. Smith DB, Bukh J, Kuiken C, Muerhoff AS, Rice CM, Stapleton JT, SimmondsP (2014) Expanded classification of hepatitis C virus into 7 genotypes and 67 subtypes: updated criteria and genotype assignment web resource. *Hepatology* 59: 318-327.
5. Takahashi K, Asabe S, Wieland S, Garaigorta U, Gastaminza P, Isogawa M., Chisari FV (2010) Plasmacytoid dendritic cells sense hepatitis C virus-infected cells, produce interferon, and inhibit infection. *P Natl Acad Sci* 107: 7431-7436.
6. Hajarizadeh B, Grebely J, Dore GJ (2013) Epidemiology and natural history of HCV infection. *Nat Rev Gastroenterol Hepatol* 10: 553.
7. Gane E J, Stedman CA, Hyland RH, Ding X, Svarovskaia E, Symonds WT, Berrey MM (2013) Nucleotide polymerase inhibitor sofosbuvir plus Ribavirin for hepatitis C. *N Engl J Med* 368: 34-44.
8. Soriano V, Vispo E, de Mendoza C, Labarga P, Fernandez-Montero JV, Poveda E, Barreiro P (2013) Hepatitis C therapy with HCV NS5B polymerase inhibitors. *Expert Opin Pharmacother* 14: 1161-1170.
9. Wendt A, Adhoute X, Castellani P, Oules V, Ansaldo C, Benali S, Bourlière M (2014) Chronic hepatitis C: future treatment. *Clin Pharmacol* 6: 1-17.
10. Ruane PJ, Ain D, Stryker R, Meshrekey R, Soliman M, Wolfe PR, Massetto B (2015) Sofosbuvir plus Ribavirin for the treatment of chronic genotype 4 hepatitis C virus infection in patients of Egyptian ancestry. *J Hepatol* 62: 1040-1046.
11. Gentile I, Borgia FR, Buonomo A, Castaldo G, Borgia G (2013) A novel promising therapeutic option against hepatitis C virus: an oral nucleotide NS5B polymerase inhibitor sofosbuvir. *Curr Med Chem* 20: 3733-3742.
12. Meissner EG, Bon D, Prokunina OL, Tang W, Masur H, O'Brien TR, Osinusi A (2013) IFNL4-ΔG genotype is associated with slower viral clearance in hepatitis C, genotype-1 patients treated with sofosbuvir and Ribavirin. *J Infect Dis* 209: 1700-1704.
13. Susser S, Herrmann E, Lange C, Hamdi N, Müller T, Berg T, Sarrazin C (2014) Predictive value of interferon-lambda gene polymorphisms for treatment response in chronic hepatitis C. *PLoS One* 9: e112592.
14. Fonseca JE, Santos M J, Canhao H, Choy E (2009) Interleukin-6 as a key player in systemic inflammation and joint destruction. *Autoimmun Rev* 8: 538-542.
15. Boltjes A, Movita D, Boonstra A, Woltman AM (2014) The role of Kupffer cells in hepatitis B and hepatitis C virus infections. *J Hepatol* 61: 660-671.

16. Arnaud C, Burger F, Steffens S, Veillard NR, Nguyen TH, Trono D, Mach F (2005) Statins reduce interleukin-6-induced C-reactive protein in human hepatocytes: new evidence for direct antiinflammatory effects of statins. *Arterioscler Thromb Vasc Biol* 25: 1231-1236.
17. Samson M, Audia S, Janikashvili N, Ciudad M, Trad M, Fraszczak J, Bonnotte B (2012) Brief report: inhibition of interleukin-6 function corrects Th17/Treg cell imbalance in patients with rheumatoid arthritis. *Arthritis Rheum* 64: 2499-2503.
18. Naugler WE, Sakurai T, Kim S, Maeda S, Kim K, Elsharkawy AM, Karin M (2007) Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. *Science* 317: 121-124.
19. Machida K, Cheng KT, Sung VMH, Levine AM, Fong S, Lai MM (2006) Hepatitis C virus induces toll-like receptor 4 expression, leading to enhanced production of beta interferon and interleukin-6. *J Virol* 80: 866-874.
20. Mohamed AA, El-Toukhy N, Reyad EM, Eldesoky NAR (2017) Serum interleukin-6 concentration Associated with Response to Therapy for Chronic Hepatitis C Patients. *J Gastroen Hepatol* 6: 2405-2410.
21. Chan YH (2003) Biostatistics 102: quantitative data-parametric & non-parametric tests. *Singap Med J* 44: 391-396
22. Chan YH (2003) Biostatistics 103: qualitative data-tests of independence. *Singap Med J* 44: 498-503.
23. Gomaa A, Allam N, Elsharkway A, El Kassas M, Waked I (2017) Hepatitis C infection in Egypt: prevalence, impact and management strategies. *Hepat Med* 9: 17-25.
24. Kamal SM, El Kamary SS, Shardell MD, Hashem M, Ahmed IN, Muhammadi M, Moniem M (2007) Pegylated interferon alpha-2b plus Ribavirin in patients with genotype 4 chronic hepatitis C: the role of rapid and early virological response. *Hepatology* 46: 1732-1740.
25. Bennett JE, Dolin R. and Blaser MJ (2015) Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases. Philadelphia: Elsevier. 563-572.
26. Cha A, Budovich A (2014) Sofosbuvir: a new oral once-daily agent for the treatment of hepatitis C virus infection. *P T* 39: 345-352.
27. Chen J, Florian J, Carter W, Fleischer RD, Hammerstrom TS, Jadhav PR., Birnkrant D (2013) Earlier sustained virological response end points for regulatory approval and dose selection of hepatitis C therapies. *Gastroenterology* 144: 1450-1455.
28. MartinotPeignoux M, Stern C, Maylin S, Ripault MP, Boyer N, Leclere L, Moucari R (2010) Twelve weeks posttreatment follow up is as relevant as 24 weeks to determine the sustained virological response in patients with hepatitis C virus receiving pegylated interferon and Ribavirin. *Hepatology* 51: 1122-1126.
29. Thorlund K, Druyts E, Mills EJ (2014) SVR12 is higher than SVR24 in treatment-naive hepatitis C genotype 1 patients treated with peginterferon plus Ribavirin. *Clin epidemiol*: 49-58
30. Satsangi S, Mehta M, Duseja A, Taneja S, Dhiman RK, Chawla Y (2017) Dual treatment with sofosbuvir plus Ribavirin is as effective as triple therapy with pegylated interferon plus sofosbuvir plus Ribavirin in predominant genotype 3 patients with chronic hepatitis C. *J Gastroen Hepatol* 32: 859-863.
31. Stättermayer AF, Strassl R, Maieron A, Rutter K, Stauber R, Strasser M, Gschwantler M. (2014) Polymorphisms of interferon- λ 4 and IL28B—effects on treatment response to interferon/Ribavirin in patients with chronic hepatitis C. *Aliment Pharmacol Ther* 39: 104-111.
32. El Salam FMA, El Toukhy NE, Mohamed AA, Nekola HA (2017) Serum interleukin-6 concentration and association with response to hepatitis C virus therapy for chronic hepatitis C patients. *Benha Med J* 34: 59-65.
33. El Serafi TI, Awad MM, Eldeen LAT, El Serafi AT, Husin M (2013) Effect of interleukin-6 and insulin resistance on early virological response of Egyptian chronic hepatitis C patients to combined pegylated interferon plus Ribavirin therapy. *Egy Liver J* 3: 21-27.
34. Feldmann G, Nischalke HD, Nattermann J, Banas B, Berg T, Teschendorf C, Sauerbruch T (2006) Induction of interleukin-6 by hepatitis C virus core protein in hepatitis C-associated mixed cryoglobulinemia and B-cell non-Hodgkin's lymphoma. *Clin Cancer Res* 12: 4491-4498.
35. Cotler SJ, Reddy KR, McCone J, Wolfe DL, Liu A, Craft TR, Ganger DR (2001) An analysis of acute changes in interleukin-6 levels after treatment of hepatitis C with consensus interferon. *J Interf Cytok Res* 21: 1011-1019.
36. Nattermann J, Vogel M, Berg T, Danta M, Axel B, Mayr C, Lutz T (2007). Effect of the interleukin-6 C174G gene polymorphism on treatment of acute and chronic hepatitis C in human immunodeficiency virus coinfecting patients. *Hepatology* 46: 1016-1025.
37. Faisal A, Zytoon AA, Gad Allah A, Dawood A (2013) Predictors of early virological response of viral hepatitis C to combination therapy with pegylated interferon plus Ribavirin. *Am J Clin Med Res* 1: 54-60.
38. Villani R, Facciorusso A, Bellanti F, Tamborra R, Piscazzi A, Landriscina M, Serviddio G (2016) DAAs rapidly reduce inflammation but increase serum VEGF level: a rationale for tumor risk during anti-HCV treatment. *PloS One* 11: e0167934.
39. Tarragô AM, da Costa AG, Pimentel JPD, Gomes STM, Freitas FB, Lalwani P, Sadahiro A (2014) Combined impact of hepatitis C virus genotype 1 and interleukin-6 and tumor necrosis factor- α polymorphisms on serum levels of pro-inflammatory cytokines in Brazilian HCV-infected patients. *Hum Immunol* 75: 1075-1083.
40. Saraiva GN, Rosário NFD, Medeiros T, Leite PEC, Lacerda GDS, Andrade TGD, Silva AA (2018) Restoring Inflammatory Mediator Balance after Sofosbuvir-Induced Viral Clearance in Patients with Chronic Hepatitis C. *Mediators Inflamm* 2018: 8578051.

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Conflict of interests: No conflict of interests is declared.