# Case Report

# Sequential *Mycoplasma pneumoniae* pneumonia and *Chromobacterium violaceum* skin abscess in a pediatric patient

Weizhen Guo<sup>1\*</sup>, Iris Wai Sum Li<sup>2,4\*</sup>, Xi Li<sup>1</sup>, Hua Xu<sup>3</sup>, Dongrong Lu<sup>1</sup>, Yue Liu<sup>1</sup>, Jiukai Chen<sup>1</sup>, Chris Ka Pun Mok<sup>4</sup>, Yingchun Zhou<sup>1</sup>

<sup>1</sup> Department of Clinical Laboratory, The First Affiliated Hospital of Guangzhou University of Chinese Medicine, Guangzhou, China

<sup>2</sup> Queen Mary Hospital, Hospital Authority, Hong Kong Special Administrative Region, Guangzhou, China

<sup>3</sup> Department of Pediatrics, The First Affiliated Hospital of Guangzhou University of Chinese Medicine, Guangzhou, China

<sup>4</sup> HKU-Pasteur Research Pole, School of Public Health, Li Ka Shing Faculty of Medicine, The University of Hong Kong, Hong Kong Special Administrative Region, China

\* Both authors contributed equally to this work.

#### Abstract

*Mycoplasma pneumoniae* is a common atypical respiratory pathogen causing community-acquired pneumonia in children. Co-infection with other respiratory viruses is common in pediatric patients but super-infection with bacteria other than *Streptococcus pneumoniae* and *Haemophilus influenzae* is rare. The first case of *Chromobacterium violaceum* infection incubated during and manifested after pneumonia caused by *Mycoplasma pneumoniae* in a 12-month old girl without any known history of immunodeficiency is here reported. The patient developed fever with redness and swelling over the middle phalanx of the right hand index finger which progressed to the formation of skin abscess. Following a course of intravenous meropenem and surgical drainage of the skin abscess, the patient fully recovered and was discharged.

Key words: Chromobacterium violaceum; Mycoplasma pneumoniae; abscess; pediatrics

J Infect Dev Ctries 2017; 11(8):656-661. doi:10.3855/jidc.8878

(Received 30 May 2016 - Accepted 26 July 2016)

Copyright © 2017 Guo *et al.* This is an open-access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

#### Introduction

Mycoplasma pneumoniae is a common atypical respiratory pathogen and is responsible for 20% to 40% of community-acquired pneumonia (CAP) cases in children and adults [1,2]. It accounts for up to 11% of the upper and lower respiratory tract infection (RTI) cases in children  $\leq 14$  years old, and around 21% and 6% in children aged  $\geq$ 5 and  $\leq$ 1 respectively [3]. Extrapulmonary manifestations have been shown occurring in up to 25% of Mycoplasma pneumoniae infected patients, more commonly in pediatric patients [4]. Coinfection with other respiratory viruses is common and occurs in up to 15% of Mycoplasma pneumoniae infected pediatric patients [2,3]. Co-infection or superinfection with bacteria other than Streptococcus pneumoniae and Haemophilus influenzae has been rarely reported [5,6]. Human infections caused by Chromobacterium violaceum have been considered as rare but with a high case-fatality rate as concluded from several case reports [7].

Chromobacterium violaceum belongs to the ß-Proteobacteria class under the family Neisseriaceae, and is a facultative anaerobe. It is a large gram-negative bacillus sized  $0.5 - 1 \mu m$  in length and 2 -  $3\mu m$  in width, and motile with a polar flagellum and one or two lateral flagella. It is a free-living organism, ubiquitously and saprophytically inhabiting in the soil and water in tropical and sub-tropical regions. Isolates from natural habitats are capable of adapting and surviving in diverse adverse environments and ecosystems [8-11]. Chromobacterium violaceum was first described as a potential pathogen in water buffalos in the Philippines by Wooley in 1905 and in humans in Malaysia by Lesslar in 1927 Several cases [10,12]. of Chromobacterium violaceum human infections have since been reported despite of the bacterium being considered as of low infectivity and usually not as a human pathogen [9,10]. The type III secretory system (TTSS) components of Chromobacterium violaceum encoded by open reading frames (ORFs), as revealed by

complete genome sequencing, allow secretions of effector molecules into the host cells leading to cytoskeletal rearrangement, and have been found to differ from those TTSS in other gram-negative pathogenic bacteria. The lack of ORFs responsible for invasions such as invI and invH, and the secretion of tyrosin phosphatase SptP in *Chromobacterium violaceum* may explain its opportunistic pathogenicity [9,13].

The first case of *Chromobacterium violaceum* skin abscess developed after *Mycoplasma pneumoniae* pneumonia in a pediatric patient without any known history of immunodeficiency is here reported.

#### Case Report

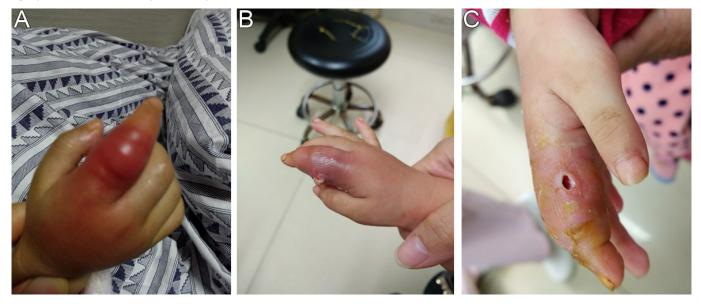
A 12-month old girl was admitted in November 2015 for recurrent fever without localized symptoms.

She presented in our hospital 3 days after discharge from another hospital where she had been hospitalized for community-acquired pneumonia due to Mycoplasma pneumoniae, presented with a 5-day history of on and off fever with peak temperature of 40°C, running nose, non-productive cough, and bronchopneumonia upon chest X-ray examination. Clinical improvement had been observed after initial empirical antibiotic treatment directed towards the most common respiratory pathogens involved in typical and atypical pneumonia. Her serum had been subsequently tested positive to Mycoplasma pneumoniae IgM antibodies, while nasopharyngeal aspirates and sputum samples were negative for respiratory viruses and bacterial pathogens, respectively.

At admission to our hospital, she had fever and nonproductive cough, without dyspnea, and no skin rash or other obvious symptoms suggestive of extra-pulmonary manifestations of the recent Mycoplasma pneumoniae infection. The patient was hemodynamically stable and her physical examination was unremarkable, except signs of pharyngitis and enlarged tonsils. Laboratory tests showed white blood cell count of  $40 \times 10^9$  cells/L with 76% neutrophils and 8% lymphocytes, hemoglobin 10.4g/dL, platelets  $359 \times 10^{9}/L$ , and Creactive protein (CRP) at 195.5 mg/L. Her serum was still positive for Mycoplasma pneumoniae IgM antibodies and negative for Legionella pneumophila serogroups 1-7, Chlamydia and Rickettsia species, Adenoviruses, Influenza viruses A & B, Respiratory syncytial virus and Parainfluenza viruses. She was given a 3-day empirical cefoperazone-sulbactam treatment for the management of her non-responding pneumonia, however her fever persisted with elevated white blood cell count of  $29 \times 10^9$  cells/L and CRP at 127 mg/L.

On day 6 after admission in our hospital, she developed redness and swelling over the middle phalanx of the right index finger and hand (Figure 1a and b). X-ray of the right hand showed soft tissue swelling of the index finger without bone erosion. The swelling progressed to abscess with pus discharge requiring surgical drainage which was performed on day 11 after admission. The pus was cultured on blood agar at 37°C with 5% CO2 and after 24 hours of incubation, a growth of round, smooth and convex colonies of 1 to 2mm diameter with β-haemolysis was

Figure 1. A and B: Skin abscess of right index finger with pus discharge and surrounding cellulitis in the right hand. C: Wound healing in progress at 4 days after surgical drainage of the abscess due to *Chromobacterium violaceum*.



observed, and presence of violet pigments was also noted when grown on Mueller Hinton agar (Figure 2) for antibiotic susceptibility test. Gram staining showed gram-negative bacilli and the isolates were subsequently identified by the automated microbial identification platform VITEK2 System with biochemical reaction results compatible to Chromobacterium violaceum. The isolate was sensitive to aminoglycosides, nitrofurantoin. trimethoprim/sulfonamides, quinolones and meropenem, and resistant to ampicillin, betalactam/beta-lactamase inhibitors, first-, second-, thirdand fourth-generation cephalosporin. The antibiotic treatment was changed to intravenous meropenem and subsequently gradual resolution of the fever and the signs of inflammation of the skin lesion were observed. Over the next 4 days post-surgical drainage, she remained afebrile, the right middle finger wound was in healing without discharge or gapping (Figure 1c), the white blood cell count was normalized and was subsequently discharged home. Furthermore, subsequent investigations for HIV antibodies and neutrophil dysfunction yielded negative results.

### Discussion

This is the first case report of *Chromobacterium* violaceum skin abscess incubated during and manifested after *Mycoplasma pneumoniae* pneumonia in a pediatric patient without any known history of immunodeficiency, and patient survival after successful treatment with meropenem.

Chromobacterium violaceum is usually considered as an opportunistic pathogen in humans. Nevertheless, an increasing number of Chromobacterium violaceum infection cases have been reported in recent decades worldwide [14]. The clinical course is characterized by acute febrile illness, rapidly progressing to lifethreatening fulminant sepsis and multi-organ failure requiring critical care, and characterized by a high mortality rate (50% to 80% in days to weeks after the infection) if left untreated. The significant risk factors for mortality in patients with Chromobacterium violaceum infection include the presence of localized abscess and the use of short clinical courses of inappropriate antibiotic treatments. Other risk factors include patient's young age and immunodeficiency status [10,13]. Relapse or recurrent infection have been reported at 6.6% [14,15].

The most common presenting symptoms of human *Chromobacterium violaceum* infection are fever, sepsis, skin lesions, abdominal pain, and clinical syndrome of localized skin and soft tissue infection,

**Figure 2.** Violet pigments of *Chromobacterium violaceum* on Mueller Hinton agar, after incubation at 37°C with 5% CO2 for 24 hours.



lymphadenitis and gastroenteritis [8,10,16]. During the infection progression bacteremia and disseminated infection involving various organ systems can result in urinary tract infection, arthritis, osteomyelitis, abscesses in lung, brain and other viscera, meningitis, endocarditis, endophthalmitis, otitis media, vaginitis, haemophagocytic syndrome and multi-organ failure [8,10,16].

The routes of infection due to *Chromobacterium violaceum* are usually through direct inoculation into mucocutaneous break or ingestion of contaminated water. Most infected individuals usually report a preceding history of trauma or wound, and exposure to recreational or stagnant muddy water, or breast surgery or appendicectomy. Isolates of *Chromobacterium violaceum* have been detected in drinking water springs, pasteurized milk samples from a dairy plant, and in samples of tap water from operation theatres. In the latter case, water has been implicated in the nosocomial transmission of *Chromobacterium* violaceum with outbreak potential and public health issues [10,17-19].

The clinical presentation of fever and skin abscess in our patient was concordant with the most commonly reported manifestation of *Chromobacterium violaceum* infection. Fifty percent of the infected patients have localized abscess [14]. Due to the prompt diagnosis of the infection, the adequate surgical drainage and the use of the appropriate antibiotics, our patient survived to the infection, and progression to bacteremia and disseminated infection was haulted despite our patient's young age.

The identification of Chromobacterium violaceum as the co-existing pathogen in our patient was prompted by the presence of non-diffusible violet pigmented colonies on blood agar, which facilitated diagnosis and treatment. About 91% of Chromobacterium violaceum strains give rise to pigmented colonies. However, there are infectious nonpigmented strains and there are also reports of co-infection with pigmented and nonpigmented strains making diagnosis even more difficult [20,21]. The organism produces a natural antibiotic called violacein encoded by a cluster of 4 genes, vioABCD, within a single operon. Its anticancerous and anti-bacterial activities would have potential implications in future clinical therapeutic or biotechnological applications with industrial benefits [13,22,23].

Our patient did not have any immunodeficiency state, such as chronic granulomatous disease (CGD), neutrophil dysfunction, HIV infection, or any other immunodeficiency, that would have predisposed her to the infection [7,10]. Moreover, she did not have any apparent history of trauma, injury, wound or exposure that would have allowed direct inoculation of organisms to the skin so as to establish the infection. Nevertheless, it is believed that Chromobacterium violaceum infection in our patient was incubated during Mycoplasma pneumoniae infection and manifested sequentially. Mycoplasma pneumoniae is well known as a great mimicker, therefore almost any organ system can be affected. The most common extra-pulmonary manifestation is exanthematous skin rash with or without mucosal lesions [4,24]. *Mycoplasma* pneumoniae infection in our patient was evidenced by the elevated Mycoplasma pneumoniae-specific IgM antibodies, which typically appear within 1 week after the infection, with a more prominent response in children than in adults, and around 2 weeks before detectable IgG antibodies [4]. This preceding episode of acute infection of Mycoplasma pneumoniae in our patient may have caused unnoticed cutaneous lesions as an extra-pulmonary manifestation of the Mycoplasma pneumoniae infection. Even a minute skin crack or break in our patient's skin may have resulted in reduced local immunity, which in turn may have facilitated the subsequent entrance of Chromobacterium violaceum pathogens leading to infection as well as skin abscess.

Moreover, *Mycoplasma pneumoniae* has been shown to induce transient depression of T-lymphocyte function and depletion of CD4 T-cells, and a transient anergy state during the acute phase of infection [25]. Temporary suppression of the immune system has also been shown in pediatric patients acutely infected by Mycoplasma pneumoniae [25]. In addition, blood lymphocytes from patients in acute phase of Mycoplasma pneumoniae infection have been shown to have a decreased response to purified protein derivatives in vitro [26]. Hence, it has been suggested that Mycoplasma pneumoniae infection causes transient depression of cell-mediated immunity [26]. It appears possible that the depressed cell-mediated immunity during the acute Mycoplasma pneumoniae infection predisposed our patient to the development of Chrombobacterium violaceum infection which manifested as skin abscess. The interval of 3 days between the resolution of the Mycoplasma pneumoniae infection symptoms and the onset of the Chromobacterium violaceum infection symptoms in our patient is compatible with the median incubation period reported for Chromobacterium violaceum (3 days median, range 1 to 90 days) [14]. It is also concordant with the sequential manifestation of *Mycoplasma* pneumoniae pneumonia and Chromobacterium violaceum skin abscess in our patient. Co-infection of Mycoplasma pneumoniae with respiratory viruses has been previously reported, but coinfection or superinfection with bacteria other than Streptococcus pneumoniae or Haemophilus influenzae have rarely been reported. There is no previous report of sequential Mycoplasma pneumoniae infection followed by Chromobacterium violaceum skin abscess a pediatric patient without any known in immunodeficiency or any other predisposing factor.

Mycoplasma pneumoniae is a well-recognized respiratory pathogen in children and adults, infecting the upper and lower respiratory tract leading to upper respiratory tract infection, bronchitis, bronchiolitis, tracheobronchitis, community-acquired pneumonia, associated with asthmatic exacerbation. and Mycoplasma pneumoniae are primarily mucosaassociated organisms capable of intracellular existence, closely associated with and epithelial cells extracellularly. Our patient did not have apparent extrapulmonary manifestations of Mycoplasma pneumoniae infection which are mainly due to indirect effects rather than the presence of the organism in the target organs or systems. Commonly reported infections involve the central nervous system with aseptic meningitis, encephalitis, transverse myelitis, and the cardiovascular system with myocarditis, pericarditis or acute myocardial infarction. Less commonly reported manifestations involve the gastrointestinal tract with pancreatitis, cholestatic hepatitis; haematologically

resulting into hemolytic anaemia, coagulopathy, presence of cold agglutinins, vasculitis and immunologically involving kidney, musculoskeletal with arthritis, myalgia, and rhabdomyolysis [4,25].

Chromobacterium violaceum is characteristically resistant to penicillin, first- and second-generation cephalosporins, and has a variable susceptibility to third-generation cephalosporins, carbapenems and aminoglycosides, resulting from the presence of numerous ORFs associated with various drug resistance mechanisms in particular beta-lactam and multidrug resistance genes [25,26]. It is hypothesized that these genes are essential for Chromobacterium violaceum survival in competing with other bacteria in different ecosystems, because isolates from environment have demonstrated a higher level of antibiotic resistance than laboratory reference strains [27,28]. The persistent fever in our patient did not respond to cefoperazonesulbactam but improved after adequate drainage of the abscess in combination with the administration of meropenem.

In conclusion, the first case of a pediatric patient without apparent significant exposure history or immunodeficiency who survived after *Chromobacterium violaceum* skin abscess incubated during and manifested after *Mycoplasma pneumoniae* pneumonia was reported. The management of *Chromobacterium violaceum* infection requires its prompt inclusion in the differential diagnosis, and the consideration of its characteristic multidrug resistance nature and high case-fatality rate by timely and appropriate use of antibiotics.

#### References

- Zhou Z, Li X, Chen X, Luo F, Pan C, Zheng X, Tan F (2015) Macrolide-resistant *Mycoplasma pneumoniae* in adults in Zhejiang, China. Antimicrob Agents Chemother 59: 1048-1051.
- 2. Waites KB, Balish MF, Atkinson TP (2008) New insights into the pathogenesis and detection of *Mycoplasma pneumoniae* infections. Future Microbiol. 3: 635-648.
- Chen ZR, Yan YD, Wang YQ, Zhu H, Shao XJ, Xu J, Ji W (2013) Epidemiology of community-acquired *Mycoplasma Pneumoniae* respiratory tract infections among hospitalized Chinese children, including relationships with meteorological factors. Hippokratia17: 20-26.
- 4. Waites KB, Talkington DF (2004) *Mycoplasma pneumoniae* and its role as a human pathogen. Clin Microbiol Rev 17: 697-728.
- 5. Kuroki H, Morozumi M, Chiba N, Ubukata K (2004) Characterization of children with *Mycoplasma pneumoniae* infection detected by rapid polymerase chain reaction technique. J Infect Chemother 10: 65-67.
- Chiu CY, Chen CJ, Wong KS, Tsai MH, Chiu CH, Huang YC (2015) Impact of bacterial and viral coinfection on

mycoplasmal pneumonia in childhood community-acquired pneumonia. J Microbiol Immunol Infect 48: 51-56.

- Karthik R, Pancharatnam P, Balaji V (2012) Fatal *Chromobacterium violaceum* septicemia in a South Indian adult. J Infect Dev Ctries 6: 751-755. doi: 10.3855/jidc.1866.
- 8. Kumar MR (2012) *Chromobacterium violaceum*: A rare bacterium isolated from a wound over the scalp. Int J Appl Basic Med Res 2: 70-72.
- Miki T, Iguchi M, Akiba K, Hosono M, Sobue T, Danbara H, Okada N (2010) Chromobacterium pathogenicity island 1 type III secretion system is a major virulence determinant for *Chromobacterium violaceum*-induced cell death in hepatocytes. Mol Microbiol 77: 855-872.
- 10. Moore CC, Lane JE, Stephens JL (2001) Successful treatment of an infant with *Chromobacterium violaceum* sepsis Clin Infect Dis 32: E107-110.
- 11. Sivendra R, Lo HS (1975) Identification of *Chromobacterium* violaceum: pigmented and non-pigmented strains. J Gen Microbiol 90: 21-31.
- 12. Sneath PH, Whelan JP, Bhagwan Singh R, Edwards D (1953) Fatal infection by *Chromobacterium violaceum*. Lancet 265: 276-277.
- 13. Brazilian National Genome Project Consortium (2003) The complete genome sequence of *Chromobacterium violaceum* reveals remarkable and exploitable bacterial adaptability Proc Natl Acad Sci USA 100: 11660-11665.
- 14. Yang CH, Lin YH (2011) *Chromobacterium violaceum* infection: A clinical review of an important but neglected infection. J Chin med Assoc 74: 435-441.
- 15. Sureisen M, Choon SK, Tai CC (2008) Recurrent *Chromobacterium Violaceum* Infection in a Patient with Chronic Granulomatous Disease. Med J Malaysia 63: 346-347.
- Swain B, Otta S, Sahu KK, Panda K, Rout S (2014) Urinary tract infection by *Chromobacterium violaceum*. J Clin Diagn Res 8: DD01-02.
- Byamukama D, Farnleitner AH, Kansiime F, Manafi M, Burtscher M, Mach RL (2005) Contrasting occurrence of *Chromobacterium violaceumin* tropical drinking water springs of Uganda. J Water Health 3: 229-238.
- Ambily R, Beena AK, Krishna SV (2014) Detection of *Chromobacterium Violaceum* in pasteurized milk samples from an organized dairy plant in Thrissur, Kerala. IJAR 3: 420-421.
- Umadevi S, Kumar S, Stephen S, Joseph NM (2013) *Chromobacterium violaceum*: A potential nosocomial pathogen. Am J Infect Control 41: 386.
- 20. Yang CH (2011) Non-pigmented *Chromobacterium violaceum* bacteremic cellulitis after fish bite. J Microbiol Immunol Infect 44: 401-405.
- Lee J, Kim JS, Nahm CH, Choi JW, Kim J, Pai SH, Moon KH, Lee K, Chong Y (1999) Two cases of *Chromobacterium violaceum* infection after injury in a subtropical region. J Clin Microbiol 37: 2068-2070.
- 22. Choi SY, Yoon KH, Lee JI, Mitchell RJ (2015) Violacein: properties and production of a versatile bacterial pigment. Biomed Res Int 2015: 465056.
- 23. Durán N, Menck CF (2001) *Chromobacterium violaceum*: A review of pharmacological and industrial perspectives. Crit Rev Microbiol 27: 201-222.
- Lind K (1983) Manifestations and complications of Mycoplasma pneumoniae disease: a review. Yale J Biol Med 56: 461-468.

- 25. Sánchez-Vargas FM, Gómez-Duarte OG (2008) *Mycoplasma pneumoniae* an emerging extra-pulmonary pathogen. Clin Microbiol Infect 14: 105-117.
- 26. Mogensen HH, Lind K (1980) *In vitro* stimulation of blood lymphocytes from *Mycoplasma pneumoniae* infected patients with pneumonia and with disorders of the central nervous system. Acta Pathol Microbiol Scand C 88: 61-65.
- Centro Pluridisciplinar de Pesquisas Químicas (2004) Drug resistance in Chromobacterium Violaceum. Genet Mol Res 31: 134-147.
- 28. Lima-Bittencourt CI, Astolfi-Filho S, Chartone-Souza E, Santos FR, Nascimento AM (2007) Analysis of

Chromobacterium sp. Natural isolates from different Brazilian ecosystems. BMC Microbiol 7:58.

## Corresponding author

YingChun Zhou Department of Clinical Laboratory, The First Affiliated Hospital of Guangzhou University of Chinese Medicine No.16 Airport Road, Guangzhou 510405, P.R.China. Phone: 86-36591912 Email: yingchunbaby@126.com

Conflict of interests: No conflict of interests is declared.