**Brief Original Article**

**Wide spread of OXA-48-producing Enterobacteriaceae in Algerian hospitals: A four years’ study**

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**Abstract**

Introduction: The aim of this study was to investigate the presence of carbapenemase-producing Enterobacteriaceae (CPE) in Algerian hospitals and to characterize the molecular types of carbapenemases found.

Methodology: During a four years study lasting between 2012 and 2015, 81 strains of Enterobacteriaceae with reduced susceptibility to carbapenems were collected from different hospitals. Carbapenemase genes were detected by PCR. Multi locus sequence typing was used to study genetic relationships between carbapenemase-producing *Klebsiella pneumoniae* isolates.

Results: Among 56 confirmed CPE, *blaOXA-48* was detected in 98.21% of isolates. Two isolates co-expressed NDM, and a single one was only an NDM producer. The strains displayed various susceptibility patterns to antibiotics with variable levels of resistance to carbapenems. Multilocus sequence typing (MLST) revealed the presence of multiple sequence types in circulation.

Conclusions: This report highlights the wide distribution of several clones of OXA-48-producing Enterobacteriaceae in Algeria. Urgent action should be taken to avoid epidemic situations.

**Key words:** Carbapenemase; OXA-48; NDM; *Klebsiella pneumoniae*; sequence type; Algeria.


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**Introduction**

Over the last decade, carbapenem resistance among Enterobacteriaceae has been extensively observed worldwide, seriously compromising the usage of carbapenems. Carbapenems are considered the drugs of choice in treating severe infections caused by multi-drug resistant strains, especially those expressing extended-spectrum-beta-lactamases.

Carbapenemase production represents the most important mechanism that affects carbapenem activity. Among the Enterobacteriaceae, three molecular types of carbapenemases are of major clinical importance since they are frequently associated with severe nosocomial infections. These are KPC (Ambler class A), NDM (class B) and OXA-48 (class D). These enzymes are widely distributed all over the world. However, specific reservoirs are identified for each type. KPC is mainly found in USA, China, Greece, and Italy. NDM is mainly present in India and Pakistan, while OXA-48 seems to be endemic in Turkey and in a lot of countries of the Mediterranean area. It is also largely found in North Africa [1].

In Algeria, data regarding carbapenemase-producing Enterobacteriaceae (CPE) are scarce since only five isolates producing VIM-19 carbapenemase and a single strain expressing *bla OXA-48* were recovered prior to 2012 [2,3].

Here we describe the wide spread of carbapenemase-producing Enterobacteriaceae (CPE) in our country during the period between January 2012 and December 2015.

**Methodology**

**Bacterial isolates**

During the reporting period, we conducted a multicenter study that aimed to investigate the presence of CPE and to characterize carbapenemase genes present in Algerian hospitals. Twelve hospital laboratories collaborated in the study. The participants were consequently invited to send their Enterobacteriaceae isolates that showed reduced
susceptibility to ertapenem (intermediate and resistant strains according to Clinical and Laboratory Standards Institute –CLSI–breakpoints) [4].

All the isolates were gathered in the central laboratory (Laboratoire de bactériologie médicale et de surveillance de la résistance aux antibiotiques; Institut Pateur of Algeria), where their identification and antimicrobial resistance profiles were verified. The isolates were then submitted to different phenotypic and molecular testing.

**Bacterial identification and antimicrobial susceptibility testing**

Isolates were identified by using API 20E system (bioMérieux, Marcy l’Étoile, France) and antimicrobial susceptibility testing was performed by the disk diffusion method according to CLSI guidelines for most of the antibiotics [4]. Minimum Inhibitory Concentrations (MICs) for carbapenems (ertapenem, meropenem and imipenem) and tigecycline were determined by using E test strips (bioMérieux, Marcy l’Étoile, France). The interpretation of results was made according to CLSI breakpoints (for most antibiotics) and EUCAST breakpoints for tigecycline. [5]

**Phenotypic detection of carbapenemases and extended spectrum beta-lactamases (ESBLs)**

Phenotypic study of different carbapenemases included a modified Hodge test (MHT), EDTA and temocillin assays [4,6–7]. ESBL production was studied using the double disk method by testing both cefotaxime ceftazidime, aztreonam and cefepime alone and in combination with clavulanic acid. A > 5 mm increase in a zone diameter for either antimicrobial agent tested in combination with clavulanic acid versus its zone when tested alone confirms an ESBL-producing strain.

**Molecular analysis of carbapenemase genes**

The main carbapenemase encoding genes (bla IMP, bla VIM, bla OXA-48, bla NDM and bla KPC) were investigated by polymerase chain reaction (PCR) using previously designed primers [8], followed by sequencing analysis (Sanger’s technique) in order to determine the exact variant of the genes. Briefly, amplicons were purified using the PureLink Quick PCR purification kit (Invitrogen, Applied Biosystems, Vilnius, Lithuania) according to the manufacturer’s instructions and sequenced using the BigDye Terminator ready reaction mix v3.1 (Perkin-Elmer Applied Biosystems, Austin, TX, USA), following the manufacturer’s instructions. The forward and reverse sequences were aligned using the CLC WorkBench7.0 software. The type of each identified gene was obtained by submitting aligned DNA sequences in the BLAST (https://blast.ncbi.nlm.nih.gov/Blast.cgi).

**Typing**

The genetic relationship between the 15 different *K. pneumoniae* isolates was determined by MLST using internal fragments of seven housekeeping genes (gap A, inf B, mdh, pgi, phoE, rpoB and tonB) according to the Diancourt et al. method (amplification and sequencing) [9]. The generated DNA sequences were deposited in the MLST database (http://bigdb.web.pasteur.fr/klebsiella/klebsiella.html) to obtain allelic profiles and sequence type (ST) designations.

The minimum spanning tree of the 15 analyzed strains was built using BioNumerics version 7.5 software. In order to assess a clonal analysis, a single isolate for each ST identified in this work, as well as STs available in the MLST database at the time of the study, were included using the eBURST version 3 program (www.mlst.net). A clonal complex or eBURST group was defined as a cluster of isolates sharing six of seven alleles.

**Results**

**Bacterial isolates**

Of the 81 isolates received during the study period, 56 (69, 13%) were carbapenemase producers. CPE were recovered from 54 patients hospitalized in 11 different hospitals located in the cities of Algiers, Blida (west of Algiers), Tizi Ouzou (east of Algiers), Oran (west Algeria) and Constantine (east Algeria). Strains were associated with different clinical infections including urinary tract infections (n = 17), wound infections (n = 13), pneumonia (n = 11), bacteremia (n = 8), catheter sepsis (n = 3), intra-abdominal infections (n = 2), vaginal infections (n = 2). Three other strains were found in fecal samples belonging to asymptomatic patients. Among the carbapenemase producing isolates, *K. pneumoniae* represented the most prevalent species, followed by *E. coli* and *E. cloacae* with 83.93% (n = 47), 8.93% (n = 5) and 7.14% (n = 4) of cases respectively.

**Antimicrobial susceptibility**

All carbapenemase-producing isolates were multi-resistant but displayed various antimicrobial susceptibility patterns. Seventy-three percent (n = 41) of the tested isolates were resistant to cefotaxime, 62.5% (n = 35) to ceftazidime and 60.71% to aztreonam.
Regarding the carbapenems, 62.5% (n = 35) of the strains remained susceptible to imipenem (MIC range 0.25-1 mg/L) and 53.57% (n = 30) to meropenem (MIC range 0.19-1 mg/L). On the other hand, resistance to ertapenem was observed in 98.21% (n = 55) of cases with MIC range from 1.5 to 32 mg/L and MIC 90 = 32 mg/L (Table 1 and 2). Among aminoglycosides, amikacin and netilmicin showed the highest activities with 76.78% (n = 43) and 69.64% (n = 39) of sensitive strains respectively, while gentamicin and tobramycin showed lower percentages of sensitivity: 35.71% (n = 20) and 39.28% (n = 22) respectively. Levofoxacin was the most active quinolone with a sensitivity rate of 76.78% (n = 43) while 53.57% (n = 30) of isolates were sensitive to ciprofloxacin and 48.41% (n = 27) to nalidixic acid. Thirty-eight (67.85%) isolates were resistant to cotrimoxazole, while chloramphenicol remained active against 76.78% (n = 43) of the tested strains. Tigecycline was the most active drug with 94.64% (n = 53) of sensitive isolates, MIC range from 0.125 to 4 mg/L and MIC 90 = 2 mg/L (Table 1 and 2). Co-expression of ESBL was noticed in 33 (58.92%) isolates.

**Molecular detection**

The molecular identification showed that OXA-48 is by far the most frequent carbapenemase among the Enterobacteriaceae isolates in Algeria (98, 21%, N = 55). Also, NDM were found in 3 isolates including two co-expressing bla_{OXA-48}. Sequencing identified the presence of the allele OXA-48 for 4 strains and the variant NDM-1 for two strains.

**Molecular epidemiology**

Multilocus sequence typing analysis allowed us to identify 11 different sequence types (STs) among the 15 analyzed *K. pneumoniae* isolates. Sequence type ST147 (n = 3) was mainly found in the hospital central de l’armée of Algiers and the university hospital of Constantine. It was linked twice to the co-production of NDM and OXA-48 carbapenemase. Four different other STs; ST101 (n = 2), ST14 (n = 2), ST336 (n = 1) and ST1412 (n = 1) were also identified in different wards of the hospital central de l’armée of Algiers (intensive care, surgery and burns). Sequence types ST13 (n = 1), ST485 (n = 1), ST870 (n = 1) and ST1593 (n = 1) were linked to strains isolated from 4 different hospitals of Algiers region, while ST348 (n = 1) was detected in university hospital of Blida and ST405 (n = 1) in the military hospital of Oran.

In summary, different STs were identified and were not linked to a geographical origin (hospital or area) (Figure 1).

Clonal analysis classified isolates into two clonal complexes designated by eBurst groups. Ten strains

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**Table 1. Antimicrobial susceptibility patterns of CPE isolates.**

<table>
<thead>
<tr>
<th>Antibiotics</th>
<th>Total number of isolates</th>
<th>Resistant N (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cefotaxime</td>
<td>56</td>
<td>41 (73)</td>
</tr>
<tr>
<td>Ceftazidime</td>
<td>56</td>
<td>35 (62.5)</td>
</tr>
<tr>
<td>Aztreonam</td>
<td>56</td>
<td>34 (60.71)</td>
</tr>
<tr>
<td>Ertapenem</td>
<td>56</td>
<td>55 (98.21)</td>
</tr>
<tr>
<td>Meropenem</td>
<td>56</td>
<td>26 (46.43)</td>
</tr>
<tr>
<td>Imipenem</td>
<td>56</td>
<td>21 (37.5)</td>
</tr>
<tr>
<td>Gentamicin</td>
<td>56</td>
<td>36 (64.28)</td>
</tr>
<tr>
<td>Amikacin</td>
<td>56</td>
<td>13 (23.21)</td>
</tr>
<tr>
<td>Tobramycin</td>
<td>56</td>
<td>34 (60.71)</td>
</tr>
<tr>
<td>Netilmicin</td>
<td>56</td>
<td>17 (30.35)</td>
</tr>
<tr>
<td>Nalidixic acid</td>
<td>56</td>
<td>29 (51.78)</td>
</tr>
<tr>
<td>Ciprofloxacin</td>
<td>56</td>
<td>26 (46.43)</td>
</tr>
<tr>
<td>Levofoxacin</td>
<td>56</td>
<td>13 (23.21)</td>
</tr>
<tr>
<td>Cotrimoxazole</td>
<td>56</td>
<td>38 (67.85)</td>
</tr>
<tr>
<td>Chloramphenicol</td>
<td>56</td>
<td>13 (23.21)</td>
</tr>
<tr>
<td>Tigecycline</td>
<td>56</td>
<td>3 (5.36)</td>
</tr>
</tbody>
</table>

**Table 2. MICs for ertapenem, meropenem, imipenem and tigecycline of CPE isolates.**

<table>
<thead>
<tr>
<th>Antibiotics</th>
<th>MIC range (µg/mL)</th>
<th>MIC50 (µg/mL)</th>
<th>MIC90 (µg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ertapenem</td>
<td>1.5-32</td>
<td>32</td>
<td>32</td>
</tr>
<tr>
<td>Meropenem</td>
<td>0.19-32</td>
<td>3</td>
<td>32</td>
</tr>
<tr>
<td>Imipenem</td>
<td>0.25-32</td>
<td>1.5</td>
<td>32</td>
</tr>
<tr>
<td>Tigecycline</td>
<td>0.125-4</td>
<td>1</td>
<td>2</td>
</tr>
</tbody>
</table>
assigned to different STs clustered in the eBurst group I, while the three isolates belonging to ST147 were part of eBurst group III. Two strains belonged to ST 1593 and ST348 respectively were singleton (Table 3).

Discussion
Since the inclusion of ertapenem in the susceptibility testing in our country from 2012, we are facing more and more clinical situations of isolating strains with decreased susceptibility to carbapenems in Enterobacteriaceae, as noticed throughout the world. In Algeria, the prevalence of resistance to third generation cephalosporins among nosocomial K. pneumoniae isolates reached 58.93% in 2015 while carbapenem resistance was around 2.33% (data of the Algerien Antimicrobial Resistant Network http://www.sante.dz/aarm/). This latter digit is likely to increase in the future in view of the strong pressure exerted by carbapenems for the treatment of serious infections due to cephalosporin-resistant enterobacteriaceae.

The data presented here, based on a 4-years survey, provide the first insight into the widespread occurrence of CPE in Algeria where carbapenemase production seems to be the main mechanism causing carbapenem resistance.

It is important to remember that carbapenem resistance in enterobacteriaceae can also be due to the combination of impermeability and production of extended-spectrum beta-lactamase (ESBL) or an over-expressed AmpC enzyme.

The isolates found in the study were associated with a large range of nosocomial infections of variable gravity. K. pneumoniae was by far the most important species.

Table 3. Sequence type and eBurst group of the 15 CPE isolates.

<table>
<thead>
<tr>
<th>eBurst group</th>
<th>Sequence type</th>
<th>Allelic profile</th>
<th>Number of isolates</th>
<th>Carbapenemase type</th>
<th>Geographical origin</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>13</td>
<td>2-3-1-1-10-1-19</td>
<td>1</td>
<td>OXA-48</td>
<td>EHS El Kettar, Algiers</td>
</tr>
<tr>
<td>1</td>
<td>485</td>
<td>2-1-1-1-7-1-12</td>
<td>1</td>
<td>OXA-48</td>
<td>EHS Zemiri, Algiers</td>
</tr>
<tr>
<td>1</td>
<td>336</td>
<td>2-1-1-1-72-4-4</td>
<td>1</td>
<td>OXA-48</td>
<td>HCA, Algiers</td>
</tr>
<tr>
<td>1</td>
<td>1412</td>
<td>2-5-1-1-4-1-18</td>
<td>1</td>
<td>OXA-48</td>
<td>HCA, Algiers</td>
</tr>
<tr>
<td>1</td>
<td>14</td>
<td>1-6-1-1-1-1-1</td>
<td>2</td>
<td>OXA-48</td>
<td>HCA, Algiers</td>
</tr>
<tr>
<td>1</td>
<td>405</td>
<td>2-1-62-3-10-4-110</td>
<td>1</td>
<td>OXA-48</td>
<td>HCA, Algiers</td>
</tr>
<tr>
<td>1</td>
<td>870</td>
<td>2-6-1-1-147-1-31</td>
<td>1</td>
<td>OXA-48</td>
<td>HCA, Algiers</td>
</tr>
<tr>
<td>1</td>
<td>101</td>
<td>2-6-1-5-4-1-6</td>
<td>2</td>
<td>OXA-48</td>
<td>HCA, Algiers</td>
</tr>
<tr>
<td>3</td>
<td>147</td>
<td>3-4-6-1-7-4-38</td>
<td>3</td>
<td>OXA-48+NDMOXA-48</td>
<td>HCA, Algiers/CHU Constantine, HCA, Algiers</td>
</tr>
<tr>
<td>1593</td>
<td>4-1-11-1-9-4-59</td>
<td>1</td>
<td>OXA-48</td>
<td>HCA, Algiers/CHU Blida, Blida</td>
<td></td>
</tr>
<tr>
<td>348</td>
<td>2-1-20-1-12-15-16</td>
<td>1</td>
<td>OXA-48</td>
<td>HCA, Algiers/CHU Blida, Blida</td>
<td></td>
</tr>
</tbody>
</table>

EHS: Etablissement Hospitalier Universitaire; HCA: Hôpital Central de l’Armée; CHU: Centre Hospitalo Universitaire; HMRU: Hôpital Militaire Régional Universitaire.

Each circle corresponds to an ST. The area of each circle corresponds to the number of isolates. Each ST is color coded according to its corresponding geographical origin. The relationships between strains are indicated by the connections between the isolates, and the numbers between the circles indicate the number of allelic differences. The black lines connecting the ST pairs indicate that they differ by three alleles (thick lines), four alleles (dotted lines), and five alleles (thin lines).

Carbapenems are still used for the treatment of CPE infections in Algeria because of the lack of other therapeutic options (piperacillin-tazobactam, tigecycline and intravenous fosfomycin are unavailable). Some CPE strains coproducing ESBL found in this work exhibited high levels of resistance to carbapenems. This is consequently of great clinical concern. Colistin remains the last drug for the treatment of infections caused by CPE in Algeria. Thus, resistance to this antibiotic should absolutely be monitored.

Figure 1. Minimum spanning tree of the MLST of the 15 carbapenemase producing Klebsiella pneumoniae isolates.
Our results showed a very large prevalence of OXA-48 type carbapenemase that could be considered endemic, and a rare presence of the NDM type amongst carbapenem non sensitive Enterobacteriaceae strains. This report consequently confirms the recent observations in neighboring countries [10-13]. Regarding NDM producers (3 isolates), in the absence of travel history data of the patients and as it is not common for Algerian patients to travel to the Indian subcontinent, we cannot dismiss the presence of autochthonous cases as previously described elsewhere [14].

Muti locus sequence typing among K. pneumoniae isolates revealed the presence of the international and multi resistant clones ST147, ST101, and ST14. ST101 is a successful clone of OXA-48 type carbapenemase and it has already been found in several countries of the Mediterranean area including those surrounding Algeria (Morocco, Libya and Tunisia) [13-15]. ST147 is a clone frequently associated to the production of different types of carbapenemases around the world (KPC, VIM, NDM and OXA-48) [15,16]. ST14 is a clone found in OXA-48 producers in Turkey [15-17] and NDM producers in India and United Kingdom [18]. Two isolates belonged to ST307 and ST405. Those STs were also found in Morocco (OXA-48) [15,19] and Spain (OXA-48, OXA-244, OXA-245) [15,20]. The other clones found in this study were rarely associated to carbapenemase production. The current spread of \textit{bla}_{OXA-48} in \textit{K. pneumoniae} is not simply linked to its diffusion among successful clones, but it is mainly due to its presence on a highly-transmissible, self-conjugative Incl/M plasmid [21].

The majority of the identified STs clustered into two major clonal complexes: I (10 strains belonging to different STs) and III (3 strains belonging to ST147).

**Conclusion**

This report has documented the widespread of OXA-48 producing Enterobacteriaceae in our hospitals and the genetic diversity of circulating clones. However, the major limitation is the absence of detailed clinical and epidemiological data that can allow us to appreciate the real impact of CPE on infected patients, outcomes and efficiency of administered treatment. Also, there is an urgent need to identify CPE to ensure adapted antibiotic therapy and to implement control measures to avoid further dissemination.

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**Conflict of interests:** No conflict of interests is declared.